

EVALUATION OF CHEMICAL COMPOSITION AND *IN SACCO* RUMINAL DEGRADATION CHARACTERISTICS OF LEAVES AND FRUITS OF CONDENSED TANNIN RICH TREE SPECIES

MSc. THESIS

By

BELACHEW ZERFU

May, 2012

JIMMA UNIVERSITY

**EVALUATION OF CHEMICAL COMPOSITION AND *IN SACCO*
RUMINAL DEGRADATION CHARACTERISTICS OF LEAVES
AND FRUITS OF CONDENSED TANNIN RICH TREE SPECIES**

MSc Thesis

**Submitted to the Department of Animal Sciences, School of Graduate Studies,
Collage of Agriculture and Veterinary Medicine, Jimma University**

**In Partial Fulfillment of the Requirements for the Degree of
Master of Science in Animal Production**

By

BELACHEW ZERFU

May, 2012

JIMMA UNIVERSIT

APPROVAL SHEET

Jimma University College of Agriculture and Veterinary Medicine Graduate Studies

As thesis research advisor, we hereby certify that we have read and evaluated this thesis prepared, under our guidance by Belachew Zerfu, entitled '**Evaluation of Chemical Composition and *In Sacco* Ruminal Degradation Characteristics of Leaves and Fruits of Condensed Tannin Rich Tree Species**' and recommend that it be submitted as fulfilling the thesis requirement.

Yisehak Kechero (Ass.Prof.)

Major advisor

Signature

Date

Taye Tolemariam (PhD)

Co-advisor

Signature

Date

As a member of the Board of Examiners of the MSc. Thesis Open Defense Examination, We certify that we have read, evaluated the Thesis prepared by Belachew Zerfu and examined the candidate. We recommended that the Thesis be accepted as fulfilling the Thesis requirement for the Degree of Master of Science in Animal Production.

Chairman person

Signature

Date

Internal Examiner

Signature

External Examiner

Signature

DEDICATION

I dedicate this thesis to my family and to all people that gave me a positive inspiration to go on with my studies.

STATEMENT OF THE AUTHORS

I declare that this thesis is my work and that all sources of material used for this thesis have been appropriately acknowledged. This thesis has been submitted in partial fulfillment of the requirements for MSc. degree at Jimma University, College of Agriculture and Veterinary Medicine and is deposited at the University Library to be made available to borrowers under the rules of the library. I solemnly declare that this thesis is not submitted to any other institution anywhere for the awards of any academic degree, diploma, or certificate.

Brief quotations from this thesis are allowable without special permission provided that an accurate acknowledgment of the source is made. Requests for permission for extended quotation from or reproduction of this manuscript in whole or in part may be granted by the Dean or Coordinator of the School of Graduate Studies or Head of the Department of Animal sciences when the proposed use of material is in the interest of scholarship. In all other cases, however, permission must be obtained from the author.

Name: Belachew Zerfu

Date of submission: May, 2012

Signature: _____

Place: Jimma University

ACKNOWLEDGMENTS

First of all I would like to thank the almighty God, for giving me not only physical and psychological capability but also strength and peacefulness to continue one more step of my journey.

My special appreciations and thanks go to my major advisor, Assistant Professor Yisehak Kechero, who was fundamental to this thesis accomplishment, without his expertise, my learning would surely be not compromised. He has not only been there with all his experience, but was teaching and helping me to develop my thoughts. His guidance and support were essential for the completion of this work. He was both my adviser and friend, through his philosophy, continued visit, constructive advice and guidance I learned many important things that helped me grow and contributed to my personal life. I would also like to thank my co-advisor, Dr. Taye Tolemariam, for his scientific support, guidance, and kindness and the time he took to correct this paper.

My thanks are owed to my families, especially my brothers for their all psychological and financial supports.

I would like to extend my gratitude to Animal science Laboratory workers such as Mrs. Kasech W/Mariam and Kasech Belete, and also to Veterinary parasitology and microbiology lab workers: Mrs Mekides Tarekegn and Genet Megersa for their patience and technical support during the entire laboratory work of the research.

I am also expressing genuine words of thanks to everyone at the Holleta Agricultural research center in the Department of Animal production especially Mr. Getu Kitaw, Mesay Hailu and the others laboratory assistants.

My several friends and colleagues, too many to list here, were behind my effort and God bless them all!

BIOGRAPHICAL SKETCH

The author, Belachew Zerfu, was born on September 29, 1988 G.C at Qimbibit *Wereda* of North Shewa zone, Oromia, Ethiopia. The author attended his elementary, secondary and preparatory schools between 1995 and 2007 G.C in the same place in Sheno elementary then, later Sheno secondary and preparatory schools. After successful completion of his high school education in 2007/2008, he joined Jimma University, College of Agriculture and Veterinary Medicine (JUCAVM) and graduated with a Bachelor of Science (B.Sc) degree in Animal Sciences, on July, 2010.

Following completion of the B.Sc degree, he joined the School of Graduate Studies of Jimma University in 2011 academic year to pursue his post graduate study for the Master of Science (M.Sc) degree in Animal production.

TABLE OF CONTENTS

Contents	Page
STATEMENT OF THE AUTHORS.....	i
ACKNOWLEDGMENTS.....	ii
BIOGRAPHICAL SKETCH.....	iii
TABLE OF CONTENTS.....	iv
LIST OF TABLES.....	vi
LIST OF TABLES IN THE APPENDICES.....	vii
LIST OF ABBREVIATIONS.....	viii
ABSTRACT.....	ix
1. INTRODUCTION.....	1
2. LITERATURE REVIEW.....	4
2.1 Feed Resources in Ethiopia.....	4
2.1.1 Natural pastures.....	4
2.1.2 Crop residues.....	4
2.1.3 Agro-Industrial by-products.....	5
2.1.4 Multipurpose tree species.....	5
2.1.4.1. Chemical composition of multipurpose trees.....	6
2.1.4.2 Multipurpose tree species in ruminant feeding systems.....	6
2.2. Methods for Estimating Rumen Degradation of Feeds.....	7
2.2.1. <i>In sacco</i> method to estimate feed degradation.....	7
2.2.2. <i>In sacco</i> dry matter and organic matter degradation of multipurpose trees.....	10
2.3. Condensed Tannins.....	10
2.3.1. Effect of tannins on the utilization of browse.....	11
2.3.2. Effect of tannins on animal production.....	12
3. MATERIALS AND METHODS.....	14
3.1. Description of the Study Area.....	14
3.2. Description of the Browse Species.....	14
3.3. Sampling, Drying and Storage of Plant Materials.....	14
3.4. Chemical Analysis.....	15
3.5. <i>In Sacco</i> Degradability Measurements.....	15
3.6. Statistical Analysis.....	17

TABLE OF CONTENTS (*Continued*)

4. RESULTS AND DISCUSSION	18
4.1. The Effect of Plant Species and Forage Parts on Chemical Composition.....	18
4.2. The Effect of Plant Species and Plant Parts on <i>In Sacco</i> Ruminal Dry Matter Disappearances.....	23
4.3. <i>In Sacco</i> Dry Matter Degradability Characteristics of the Leaves and Fruits of the Plant Species	25
4.4. The Effect of Plant Species and plant parts on <i>in sacco</i> ruminal Organic Matter Disappearances.....	27
4.5. Effect of Species and Plant Parts on <i>In Sacco</i> Organic Matter Degradability	28
4.6. The Correlation between Chemical Composition and <i>In Sacco</i> Ruminal Degradation of Dry matter and Organic Matter	30
5. SUMMARY AND CONCLUSION	34
6. REFERENCES.....	36
7. APPENDICES.....	55
7.1. Appendix. Chemical Composition and <i>In Sacco</i> degradability Tables.....	56

LIST OF TABLES

Table	Page
Table 1. Comparison of the least square means for chemical compositions (%) of leaves and fruits of the plant species	20
Table 2. Least square means for <i>in sacco</i> ruminal dry matter disappearances compared for leaves and fruits of the plant species.....	24
Table 3. Comparison of the least square means for <i>in sacco</i> ruminal drymatter degradability characteristics of leaves and fruits of the plant species.....	26
Table 4. Comparison of the least square means of leaves and fruits of the tannin rich plants for <i>in sacco</i> organic matter disappearances	28
Table 5. Comparison of the least square means for <i>in sacco</i> ruminal degradability characteristics of organic matter of leaves and fruits of the plant species.....	30
Table 6. The spearman's rho correlation coefficient (r) between chemical composition and <i>in sacco</i> dry matter and organic matter disappearances for some incubation hours.....	33

LIST OF TABLES IN THE APPENDICES

Appendix Table	page
Appendix Table 1. The least square means for chemical compositions (%) of leaves and fruits compared separately for the plant species	56
Appendix Table 2. Least square means of <i>in sacco</i> ruminal dry matter disappearance (%) of leaves and fruit of the plant species	56
Appendix Table 3. <i>In sacco</i> dry matter degradability characteristics for the leaves and fruits of the plant species	57
Appendix Table 4. Least square means for <i>in sacco</i> ruminal organic matter disappearance (%) of leaves and fruits of tannin rich trees	57
Appendix Table 5. <i>In sacco</i> organic matter degradability characteristics of leaves and fruits of selected browses species	58

LIST OF ABBREVIATIONS

AIBP	Agro-Industrial By-Products
AOAC	Association of Official Analytical Chemists
ARC	Agricultural Research Council
BFT	Birds foot Trefoil
BW	Body Weight
CRs	Crop Residues
CT	Condensed Tannin
DMD	Dry Matter Disappearance
ED	Effective degradability
FAO	Food and Agriculture Organization
GGCSWE	Gilgel Ghibe Catchments of South Western Ethiopia
HARC	Holeta Agricultural Research Center
HF	Holstein Friesian
HT	Hydrolysable Tannin
IAEA	International Atomic Energy Agency
ILRI	International Livestock Research Institute
JUCAVM	Jimma University College of Agriculture and Veterinary Medicine
MPTS	Multipurpose Tree Species
NRC	National Research Council
OMD	Organic Matter Disappearance
SAS	Statistical Analysis System
TLU	Tropical Livestock Units

Evaluation of chemical composition and *in sacco* ruminal degradation characteristics of leaves and fruits of condensed tannin rich tree species

By: Belachew Zerfu

Major advisor Yisehak Kechero(Ass.Prof) and Co-advisor Taye Tolemaria(PhD)

ABSTRACT

The objectives of this study were to determine the chemical composition, in sacco rumen dry matter and organic matter degradability of leaves and fruits of indigenous tannin rich multipurpose tree species (MPTS) and also to assess the relationship between chemical composition and in sacco ruminal degradability parameters. The MPTS studied were Ekebergia capenesis, Ficus sycomorus, Maesa lanceolata and Rhus glutinosa. The leaves and fruits of the MPTS were collected from the Omo-Nada woreda of Jimma zone, southwestern Ethiopia during East African Autumn season. Chemical composition measured were dry matter (DM), crude protein (CP), crude ash (CA), ether extract (EE), crude fiber (CF), neutral detergent fiber (NDF), non-fiber carbohydrates (NFC) and condensed tannin (CT). In situ rumen degradability was measured with of three rumen fistulated Holstein Friesian-Boran cross steers in times at 0, 6, 12, 24, 48, 72 and 96 h. Ruminal in sacco OM degradabilities were estimated from residues in nylon bags. The DM and OM degradability data were fitted to the exponential equation $Y = a + b(1 - e^{-ct})$. The values for each chemical constituents was ranged from 92.43 to 96.84% DM, 5.43 to 11.49 % CA, 7.97 to 17.06 % CP, 1.57 to 31% EE, 12.20 to 27.5% CF, 5.84 to 39.30 %NFC, and 7.2 to 16.72% CT. For DM degradation, the highest and lowest values of soluble fraction (a) of leaves were recorded in M. lanceolata (14.55% DM) and E.capensis (-16.20% DM), respectively ($P < 0.001$). The highest (0.12%/h) in E.capensis leaf and lowest (0.01%/h) for R.glutinosa leaf were the values of degradation rate(c). However in OM degradation kinetics, the 'a' value was ranged from -1.02% in R.glutinosa to 10.69% in E.capensis leaves where the variation was significant for the plant species ($P < 0.001$); on the other hand, the highest and lowest 'a' value was recorded for fruits of R. glutinosa (31.84%) and E.capensis (14.22%) ($P < 0.001$). The insoluble but degradable fraction (b) was 83.35% in R. glutinosa and 45.21% in E.capensis leaves ($P < 0.001$); On the contrary, the highest and lowest values of potential degradability (PD) was recorded for F.sycomorus (89.89%) and 55.90% for E.capensis leaves ($P < 0.001$). On the other hand, the highest and lowest effective degradability(ED) value was obtained in R.glutinosa (63.97%) and E. capensis leaves ($P < 0.001$). The degradation rate constants (c) varied widely between MPTS with similar rates for E.capensis and F.sycomorus leaves. In general, it was observed that variation of plant parts lead to highly significant differences in DM and OM degradability and the degradable parameters. DM and OM degradation were highly correlated with time taken for the incubation, CP, NFC and CT concentration. The edible parts of MPTS recorded more than 60% DM and OM degradability at 24 h, which implied that they were all highly degradable in the rumen. Their incorporation into ruminant feeding systems as dry season forage supplements is therefore highly recommended.

Key words: Chemical composition; in sacco rumen degradation; fruit; leaves; multipurpose trees; condensed tannin

1. INTRODUCTION

Inadequate feed supply is a major constraint of ruminant livestock production in sub-Saharan African countries (Arigbede *et al.*, 2011). Dry season is always a critical period when most feed resources especially grasses and herbaceous forages dry out (Kanani *et al.*, 2006). Crop by-products, available during dry season, have a low nutritive value (Salem *et al.*, 2006). This obviously adds to the poor performance of ruminant livestock. Improvement of the performance of ruminants in Sub-Saharan Africa (Solomon *et al.*, 2004; Babayemi *et al.*, 2004b; Mekoya *et al.*, 2008) call for, use methods of extending the availability and quality of local feedstuffs. One potential way for increasing the quality and availability of feeds is the use of various multipurpose trees and shrubs (MPTS) (Devendra, 1989; Kaitho *et al.*, 1998; Woldemeskel *et al.*, 2001).

Leaves, twigs, stems, pods, fruits and seeds of MPTS have been used as cheap and affordable supplements for ruminant animals in herds of resource poor farmers in several regions of the World (Salem *et al.*, 2006). These morphological fractions of MPTS has been used to supplement straws to alleviate protein deficiency in fibrous feeds, reduce rumen acidosis and other health-related problems, and improve intake and productivity of animals (Owens *et al.*, 1998; Mekoya *et al.*, 2008).

A variety of MPTS are growing in the Gilgel Ghibe catchments of Southwestern Ethiopia (GGCSWE) (Yisehak *et al.*, 2010), mainly due to the suitability of the environment and the need to use them as fire wood, local construction, mulch, and shade for cash crops like coffee and spices. They replenish soil fertility, serve as of human and veterinary medicine, and also serve as environmental conservation. Currently small-holder farmers of sub-Saharan African countries in general (Aremu and Onadeko, 2008) and South West Ethiopia in particular (Yisehak and Belay, 2011; Yisehak *et al.*, 2012) are increasingly relying on various potential browse plants that can provide a green feed throughout the year which may be particularly useful as feed supplements to the typical low-quality diets.

Among MPTS growing in the GGCSWE, *Ekebergia capensis*, *Ficus sycomorus*, *Maesa lanceolata*, and *Rhus glutinosa* are widely abundant, evergreen throughout the year, and have a good biomass yield and multiple uses and highly preferred by farmers (Yisehak *et al.*, 2010).

Most browse species have an advantage of maintaining their greenness and nutritive value throughout the dry season when grasses dry up and deteriorate both in quality and quantity. Browse is generally richer in protein and minerals (Le Houeron, 1980; Kadzere, 1995; Alonso-Diaz *et al.*, 2010) and thus has the potential to be an inexpensive, locally produced protein supplement that plays an important role in the nutrition of grazing animals (Meuret *et al.*, 1990; Salem *et al.*, 2006). Unfortunately, their content of nutrient activating factor like condensed tannins (CT) varies widely and unpredictably (Babayemi *et al.*, 2004b). The use of edible parts of tree and shrub leaves by herbivores is restricted by defending or deterring mechanisms related to high CT content (Provenza *et al.*, 1995). High levels of CTs in leaves restrict the nutrient utilization and decrease voluntary food intake, nutrient digestibility and nitrogen (N) retention (Kumar and Vaithiyanathan, 1990; Silanikove *et al.*, 1996). Their effect on animals ranges from beneficial to toxicity and death (Makkar *et al.*, 2003).

The CT content of plants is affected by plant species (Ozturk *et al.*, 2006), and stage of growth, and may vary with plant part (leaf, stem, inflorescence, seed), season of growth and other specific environmental factors such as temperature, rainfall, cutting and defoliation by grazing herbivores including insects (Makkar and Singh, 1991b). The chemical composition and digestibility of forages are also influenced by plant species, plant morphological fractions, environmental factors and stage of maturity (Papachristou and Papanastasis, 1994).

The performance of herbivores when grazing depends on forage digestibility and intake (Ramirez *et al.*, 2000; Decruyenaere *et al.*, 2009; Fraser *et al.*, 2009). Moreover, forage intake is related to fibre digestibility because intake is reduced when fibre is increased in the digestive tract (Mertens, 1993). Rate of digestion provides an important measure of forage quality because intake of forages having rapid rates of digestion is greater than that for forages with slower rates of digestion but similar total digestibility (Holechek *et al.*, 1982). Since the rumen is the primary

site of digestion of forages, it is important to monitor their degradation kinetics. This can be achieved by using *in sacco* technique which is quicker and cheaper than whole animal studies.

The *in sacco* nylon bag technique, in which feed samples in nylon bags are suspended in the rumen, is widely used to estimate the rate and extent of degradation and digestion of feed in the rumen (Marinucci *et al.*, 1992; NRC, 2001). The methodology used to generate degradability values has a major influence on degradability values published (Michalet-Doreau and Cerneau, 1991). Standardization of the method is related to a large number of factors causing variation, e.g. the bag (sample dry matter, bag surface area to sample weight ratio and cloth pore size), sample preparation (particle size of sample), pre-incubation treatment, washing procedure, diet of fistulated animals, correcting for microbial contamination of residues, etc. (Nocek, 1988; Vanzant *et al.*, 1998).

Important characteristics of digestion in the rumen with regard to forages are: effective degradability, rate of digestion and the amount of digestible fibre (Singh *et al.*, 1989; Larbi *et al.*, 1997). Rumen degradation is thus regarded as a major descriptor of forage quality (Orskov and McDonald, 1979). It is useful in ranking trees and shrubs in terms of nutritive value (Larbi *et al.*, 1994) and for comparing the digestive capabilities of forages (Singh *et al.*, 1989).

However, limited information is available on the combined effects of the factors (species, and plant parts) on chemical composition and *in sacco* ruminal degradation characteristics of the tannin rich tree plants. Therefore, the present research was initiated to investigate the following specific objectives:

- (1) To evaluate the effect of browse species and plant parts on the chemical composition and *in sacco* ruminal degradation characteristics of dry matter and organic matter and
- (2) To assess the relationship between some chemical composition and *in sacco* ruminal degradability parameters.

2. LITERATURE REVIEW

2.1 Feed Resources in Ethiopia

According to Alemayehu (2003), the major livestock feed resources of Ethiopia includes natural grazing and browse, crop residues, agro-industrial by-products, and cultivated pasture and forage-crop species.

2.1.1 Natural pastures

Natural pastures include annual and perennial species of grasses, forbs and trees. Many researchers and development workers agreed that natural pasture comprises the largest feed resource in Ethiopia but estimates of the contribution of this feed resource vary greatly. Alemayehu (1998a) estimated that 80-85% of all feed comes from natural pasture while some estimates indicate the natural pasture provides 88-90 percent. This is because the quantity and quality of native pasture varies with altitude, rainfall, soil and cropping intensity. Currently, with the rapid increase of human population and increasing demand for food, grazing lands are steadily shrinking being converted to arable lands, and are restricted to areas that have little value or farming potential such as hill tops, swampy areas, roadsides and other marginal land (Ibrahim, 1999). This is particularly evident in the mixed farming highlands and mid altitudes. Communal grazing is normal and managed as a common property resource (Behnke and Schoones, 1993; Wolmer, 1997). Due to land scarcity and crop-dominated farming (Alemayehu, 2002) there has been limited spontaneous introduction of improved pasture and forages. During the Fourth Livestock Development Project, different strategies and species for pasture and forage development were selected.

2.1.2 Crop residues

Crop residues (CRs) are roughages that become available as livestock feeds after crops have been harvested. They are distinct from agricultural by-products (such as brans, oil cakes, etc), which are generated when crops are processed. Residues can usually be grouped along crop

types-cereals, grain legumes, roots and tubers, and so on (World Bank, 1989; Nordblom and Shomo, 1995). Apart from being a source of animal feed, residues are sources of building, roofing and fencing materials. They are used also as fuel and as fertilizers or as surface mulch in cropland (Van Raay and De Leeuw, 1970, 1974). Their value as feed depends on the demand from livestock owners, which varies with the overall supply and demand situation for feeds. This, in turn, depends on the density of livestock, usually expressed in tropical livestock units per square kilo meter (TLU km²) and the supply of other feed resources, in particular, forage and browse from natural vegetation (De Leeuw and Rey, 1995). The supply of CRs is a function of the proportion of land used for cropping and the amount of edible feed yields per unit of land.

2.1.3 Agro-Industrial by-products

Agro-industrial by-products (AIBP) are derived from the processing of a particular crop or animal product usually by an agricultural firm. The AIBP widely used as livestock feed in Ethiopia are materials like molasses, bagasse, oilseed cakes and maize milling by-products, and brewer's wastes. Unfortunately, there is no factory in Ethiopia producing animal by-products (e.g. meat and bone meal, bone meal, and fish meals) (Berhanu *et al.*, 2009).

2.1.4 Multipurpose tree species

Multipurpose tree species (MPTS) are trees that provide multiple benefits and serve multiple purposes. MPTS serve purposes like: soil erosion control, shade and shelter, and soil fertility improvement and provide beneficial products like: fodder and cattle feed, food, fuel wood, green manure, manure (oil cakes), timber, pulp, tannin, gum, raw material for dyes, medicines, pesticides, posts and poles. They can be grown on farm bunds, along boundaries and in degraded areas of the farm. Generally MPTS are hardy and grow well without special care in suitable agro-climatic conditions (Mekoya, 2008).

2.1.4.1 Chemical composition of multipurpose trees

The chemical composition of the MPTS shows substantial variation across species and seasons. The DM content is higher during the dry than rainy seasons and (Asiegbu and Anugwa, 1988; Anele *et al.*, 2008) and elsewhere (Topps, 1992; Ly *et al.*, 2001). Greater DM content observed during the dry season might be as a result of reduced photosynthetic activity due probably to the lower moisture levels in the soil during this period relative to the rainy season. Contrary to this, greater CP content recorded during the rainy season might be due to re-growth of new flush of leaves during this period and their lower CP content during the dry season may be largely due to the moisture stress experienced by the trees during this period. The CP content across seasons for all MPTS investigated was adequate to support meat and milk production of ruminant animals (Lamers and Khamzina, 2010). They may therefore serve as ready source of year-round cheap protein supplements for herds of resource-poor farmers in Sub-Saharan Africa and thereby help to improve the quality of feed and at the same time reduce the cost of production.

Reports from literature confirmed that ruminant animals are able to handle browse plants with tannin content below 100 mg/g DM (Gasmi-Boubaker *et al.*, 2005) although, the tolerance level may vary between animal species (Onwuka, 1992). Apart from seeds, trypsin inhibitors and phytic acid have not been reported at levels that could pose nutritional dangers to ruminant animals in most tropical browse species, thus, their analysis before feeding may become optional for farmers in Sub-Saharan Africa.

2.1.4.2. Multipurpose tree species in ruminant feeding systems

Differences in chemical composition and *in sacco* dry matter degradation characteristics have practical implications for MPTS-based agro-forestry technologies and ruminant feeding systems. Because of their high CP content and potential extent of DM and OM degradation as compared to majority of grass species, they can be a source of rapidly degradable nutrients for microbial protein synthesis in the rumen (Lamers and Khamzina, 2010). Incorporation of MPTS foliage with animal feed could increase the CP content to above 10% of DM, even at low levels of supplementation. With depleted grazing resources in the dry season, the grazing period could be drastically reduced and level of foliage intake increased to 50%. This practice can increase the

CP content of diet to 14% or above during the dry season and at the same time reduce the cost of feeding and amount of energy normally expended on trekking in search for grazing resources (Patra, 2010).

2.2. Methods for Estimating Rumen Degradation of Feeds

Currently, numerous methods involving different procedures are available for estimating degradation of feeds like, the *in vivo* methods involve markers and the *in sacco* method requires animals that are surgically modified with rumen cannulae. The *in vitro* methods require the use of rumen fluid, which is obtained from fistulated animals, to estimate either digestibility (Tilley and Terry, 1963) or gas production (Krishnamoorthy *et al.*, 2005, Menke *et al.*, 1979). But the emphasis is on the *in sacco* method to estimate feed degradation.

2.2.1. *In sacco* method to estimate feed degradation

The *in sacco* technique was first suggested by (Quin *et al.*, 1938) and it has since been used by others to estimate utilisation of either forages (Van Keuren and Heinemann, 1962) or concentrates and high-protein feeds (Mehrez and Orskov, 1977).

Interest in the technique has intensified since Mehrez and Orskov, (1977) critically assessed the factors causing variability in DM and N degradability. They concluded that as long as the bags were large enough to allow free movement of substrate within, the technique could be extremely useful as a rapid guide to determine nutrient disappearance, particularly the rate and extent of nutrient disappearance from the rumen. All modern systems of feeding ruminants (ARC, 1993, NRC, 1996, ARC, 1984) require an estimation of the amount of feed protein escaping ruminal degradation. This estimation is obtained by the *in sacco* technique, which is probably the best-known simple and reliable method to assess the degradability of DM and protein in the rumen (ARC, 1993, NRC, 1996, Thomas, 2004, Mehrez and Orskov, 1977, ARC, 1984).

The *in sacco* method requires the use of fistulated animals, which limits its routine use by the commercial laboratories. However, it is widely applied by researchers since it requires fewer measurements, is relatively less labor intensive and so is cheaper as compared with the *in vivo*

method. The *in sacco* method involves the sealing of feed samples within nylon, polyester or Dacron bags, which are then suspended in the rumen of sheep or cattle for varying periods of time, followed by determination of the DM and protein in the washed residues. The technique allows the test feed to be incubated in the ruminal environment (i.e. pH, temperature and CO₂), but unlike the normal situation the feed is not subjected to mastication and rumination. Despite its widespread use, the technique has inherent limitations that must be taken into account, particularly if comparisons of degradation among different laboratories are to be made. The sources of variations in the use of the *in sacco* method among different laboratories in terms of bag size, sample size, particle size and time (h) of incubation used by different authors. The assumption that the N leaving the bag during washing in water, at 0h of incubation, is completely degraded may not be true (Chaudhry and Webster, 2001, Mahadevan *et al.*, 1980). Extensive loss of feed material at 0h incubation times will lead to an overestimation of degradability. Although (Chaudhry, 2007, De Smet *et al.*, 1995, Djouvinov *et al.*, 1998, Kristensen *et al.*, 1982, Noziere and Michalet, 1996) present information in relation to degradable crude protein values only, it is assumed that similar variations in practice will also cause variations in DM and organic matter degradability values.

Beside microbial contamination within the bag, there are numerous other sources of errors that affect the *in sacco* DM and N disappearance from feeds. The importance of sample weight in a given bag size has been emphasized by (Bullis *et al.*, 1967) who observed reduced DM digestibility with increased weight in the bag. Van Keuren and Heineman, (1962), showed that sample weight influenced DM digestibility, at least when short incubation times were used; the difference tended to disappear with longer periods of incubation. Also, oven drying of silage samples at high temperatures was found to reduce N degradability and solubility (Lopez *et al.*, 1995) of the sample.

Additionally, Noziere and Michalet-Doreau, (2000), reported that grinding and pre-wetting underestimates degradation rates due to the increased microbial colonisation. Machine washing of residues overestimates solubles and particulate losses but it is less subjective than hand washing (Cockburn *et al.*, 1993).

Huntington and Givens, (1995), reported that bag pore size less than 15mm can reduce degradation by restricting microbial colonization and diversity and trapping fermentation gases. However, bag pore size of more than 40 mm can cause losses of soluble and undegradable particles. Furthermore, the animal effects and bag incubation sequence also contribute to the variation in results among laboratories (Nocek, 1988). The disappearance of DM is also affected by the diet fed to the host animal (Caton *et al.*, 1988). While these effects make it difficult to compare feeds for degradation across studies, DM or protein degradation of a feed is not entirely a function of the feed, but also affected by the ruminal conditions, so variation across studies is expected. The pH inside the nylon bag has been shown to be lower than that outside the bags, especially when small pore-sized bags were used (Marinucci *et al.*, 1986). The microbial population inside the bags also differed, both in composition and concentration, from that of the outside of the bag. For example, both protozoa and bacterial populations were found to be lower inside the bags (Meyer and Mackie, 1986). This could be due to the limited micro-environment that existed within the bag involving a single ingredient of smaller size with limited exposure to rumen microbes perhaps due to the bag size and its pores.

Analysis of digesta from nylon bags incubated *in vitro* showed that some nutrients escaped the nylon bags before being digested (Spencer *et al.*, 1988). The microbial attachment to feeds incubated *in sacco* is frequently not measured, though several studies have shown high levels of contamination of incubated feed with rumen microbes (Huntington and Givens, 1995). All these sources of errors increase variability of predictions of degradability among laboratories. In spite of being widely used and standardised, the application of this methodology needs to address two points: first, the fraction assumed to be completely degraded, and the DM and N disappearance during this step could simply be due to DM or N washed out of the bag; second, the microbial contamination of feeds within the bags. The first point would overestimate degradation and the second point would underestimate it (Madsen and Hvelplund, 1994).

The significance of these two factors would be important depending on the type of feed being analysed. However, the *in sacco* method is still the reference method in most countries; the reason is probably that the degradability is measured in the rumen and, therefore, from a

biological point of view it is more reliable than those of the in vitro methods (Hvelplund and Weisbjerg, 1998).

2.2.2. *In sacco* dry matter and organic matter degradation of multipurpose trees

The MPTS recorded more than 60% DM and OM degradability at 24 h which implied that they are extensively degraded in the rumen. Variation in DM and OM degradability is directly related to the proportion of structural and non-structural protein and carbohydrate fractions which in turn affects their solubility and bioavailability (Whetton *et al.*, 1997). Inter-species variations in OM and DM degradabilities could result in differential intakes of the MPTS when given as sole diets to animals (Papachristou and Papanastasis, 1994).

2.3. Condensed Tannins

Condensed tannins (CT) are secondary plant compounds found in some plant species. These plant products are polymers of flavan-3-ol (catechin) or flavan-3, 4-diol (proanthocyanidins) units linked by C-C bonds (Waghorn *et al.*, 1997; McMahon *et al.*, 2000). The CT are typically found in plant cell vacuoles (Min *et al.*, 2003), but location can vary among plant species. The CT can be found in cell walls, stems, bark, leaves, flowers, and seeds (McMahon *et al.*, 2000). In the *japonicas* variety of birdsfoot trefoil (BFT), CT were not found in leaf tissue (as in other BFT cultivars), but were observed in stems and flowers by Morris *et al.* (1993). Conversely, CT are found throughout sainfoin (*Onobrychis viciifolia* Scop.), with the highest concentration of CT being found in the leaves (Lees *et al.*, 1993). CT concentration of plants can also be influenced by maturity, temperature, and soil fertility as well as grazing (McMahon *et al.*, 2000). Tannin concentration in BFT was reported to increase with increasing plant maturity and was higher in regrowth than in spring growth in West Virginia (Cassida *et al.*, 2000).

The structure of CT can vary greatly. Monomers can be linked by C-4 and C-8 or C-4 and C-6 interflavan bonds, which alter the shape of the compound (Haslam, 1989; Barry and McNabb, 1999). The number of polymer units, and therefore the molecular weight (MW) can also vary among CT. These differences can dramatically alter the structure of CT, which can subsequently modify the nutritional effect of CT from different plant sources (McMahon *et al.*, 2000).

Condensed tannins form complexes with many compounds, including protein and carbohydrates (Barry and McNabb, 1999). Feeding forages with high concentrations of CT (10% DM or greater) has been shown to reduce feed intake, N utilization, and carbohydrate digestion (Waghorn *et al.*, 1997; Barry and McNabb, 1999). Condensed tannins from plant species such as BFT and sainfoin can bind protein by hydrogen bonding at near neutral pH (pH 6.0 to 7.0) in the rumen to form CT-protein complexes, and then dissociate and release bound protein at pH less than 3.5 in the abomasum (Barry *et al.*, 2001). This, in turn, decreases protein degradation and NH₃-N production in the rumen, which results in improved N utilization and reduces N waste excretion by the animal. This reduction in soluble protein digestion in the rumen also prevents frothy bloat, making forages containing CT safe to graze (Kendall, 1966).

2.3.1. Effect of tannins on the utilization of browse

Most browse species contain large amounts (up to 50% of the dry matter) of tannins (Leinmuller *et al.*, 1991; Cabiddu *et al.*, 1998). Tannins are complex phenolic compounds that contain sufficient hydroxyl and carboxyl groups to precipitate proteins and to bind carbohydrates under conditions that prevail in the digestive tract of mammals and birds. The negative effects of tannins on palatability and digestibility in ruminants are multiple (Kumar and Vaithyanathan, 1990). They include: (i) reduction in protein availability due to binding of food proteins and inactivation of enzymes in the digestive tract, (ii) astringency caused by the interaction of tannins with salivary protein and oral mucosa, and (iii) gut irritation and systemic toxicity. All of the aversive effects can reduce forage palatability. In general, there is an inverse relationship between tannin concentration in browse sources and voluntary feed intake by herbivores (Kumar and Vaithyanathan, 1990).

Condensed tannin contents above 3% may act as a feeding deterrent (Provenza, 1995), influence feed degradation in the rumen (Silanikove *et al.*, 1996a) and the digestibility of the whole diet (MacNaughton, 1987; Silanikove *et al.*, 1997). Hence, when tannin-rich leaves are offered as a sole feed to sheep and goats they may not provide for adequate absorbed nutrients for maintenance despite their relatively high-protein and low-fibre contents (Silanikove *et al.*, 1994, 1996a). They may also reduce animal productivity in terms of weight gain, milk yield and wool growth (Kumar and Vaithyanathan, 1990). Silanikove *et al.* (1996a) reported that tannin levels

of approximately 20% of DM drastically reduced leaf intake of *Pistacia lentiscus* and goats fed such a diet were in marked negative nitrogen balance, and lost weight very rapidly(100g/day).

Tannins can suppress intake by reducing digestibility or by causing illness. Tannins may bind to cell walls and cell solubles (Kumar and Vaithyanathan, 1990; Reed, 1995) and in the process reduce the digestion of protein and yield of energy-rich byproducts of microbial fermentation such as volatile fatty acids (Makkar *et al.*, 1995). This in turn may adversely affect preference of the feed containing the tannins (Villalba and Provenza, 1996, 1999). Tannins may also produce adverse postingestive effects that cannot be accounted for by digestion inhibition alone, primarily because they cause such rapid (within a few to 60 min) and dramatic decreases in food intake (Silanikove *et al.*, 1997b; Landau *et al.*, 2000). Silanikove *et al.* (1997b) in goats and Landau *et al.* (2000) in heifers have shown that feeding ruminants diets rich in condensed tannins was associated with lowered feed intake and shorter duration of eating bouts, mainly of the first eating bout, immediately after distribution of the diet. The data of Silanikove *et al.* (1997b) and Landau *et al.* (2000) suggest that: i) negative effects of condensed tannins derive from astringency and short-term post-ingestive malaise; ii) the increased number of eating bouts and their wider partition throughout the day are means to preserve the ruminal environment.

2.3.2. Effect of tannins on animal production

Tannin content is highest when the shrub is rapidly growing and tannin poisoning can occur when young or inexperienced livestock consume immature buds in the spring. Tannin content can also increase following summer rains. Research reported by Gomes, (1990) at the University of Arizona for turbinella oak concluded that condensed tannins are highest in the winter with immature leaves and that soluble tannins are highest in immature leaves in the summer through early fall.

Tannins, because of their protein-binding properties, are known to be strongly astringent. This astringency appears to be a major cause of reduced feed intake in mammalian herbivores. There is some controversy, however, over whether reduced food intake is a result of the nature of tannins. Singleton (1981) considers it to be unfair to consider the effects of tannins on feed

intake as considers it to be unfair to consider the effects of tannins on feed intake as consumption itself. On the other hand, Provenza *et al.* (1990) suggest that mammals may reject tannin-containing plants because they cause internal malaise. Severe growth depression can be a result of reduced feed intake, as has been shown to appear in rats and chicks when fed tannin containing diets (Fahey and Jung, 1989). When tannins complex with proteins in an animal's gut, they are believed to be responsible not only for growth depression, but also for low protein digestibility and increased faecal nitrogen concentrations (Salunkhe *et al.*, 1990). Thus, once they have been consumed, their adverse effects seem to be related to their binding of dietary protein. There is evidence that enzymatic protein as well as endogenous proteins comprise a considerable portion of excreted nitrogen when animals are fed tannins (Allredge, 1994). When endogenous proteins are lost in this manner, the animal may incur a deficiency in one or more essential amino acids.

The ability of tannins to form strong complexes with proteins is the most important aspect of their anti-nutritional effects. Tannins bind with at least four groups of proteins in the ruminant: dietary proteins, salivary proteins, endogenous enzymes and gut microbes including microbial enzymes (Hagerman and Butler, 1981). The effects on such as inhibition of feed intake and digestion by ruminants are usually ascribed to their ability to bind to proteins. The strength of the tannin-protein complexes depends on characteristics of both the tannin and protein (Haslam, 1989). HTs and CTs differ in their nutritional significance and toxic effects, but both precipitate proteins (Reed, 1995). While CTs are not readily degraded in the gut, hydrolysable tannins undergo microbial and acid hydrolysis with the release of simpler phenolics. These are absorbed and can cause toxicity. While CTs reduce forage quality, the HTs cause poisoning in animals if sufficient quantities are consumed. McSweeney *et al.* (1988) found that although sheep were sensitive to the toxicity of hydrolysable tannins present in the browse tree *Terminalia oblongata*, the digestion of N, organic matter and cell wall constituents remained unaffected. In contrast, CTs have detrimental nutritional effects such as reducing feed intake, reducing feed digestibility and increasing faecal nitrogen excretion (Reed and Soller, 1987). On the other hand, CTs can be of benefit in the prevention of bloat (Jones *et al.*, 1973), in the protection of feed protein against degradation in the rumen (Barry *et al.*, 1986) and by increasing N retention (Robbins *et al.*, 1987) under some conditions.

3. MATERIALS AND METHODS

3.1. Description of the Study Area

The chemical composition study including tannin components was carried out in animal nutrition laboratory of Jimma University, College of Agriculture and Veterinary Medicine (JUCAVM) campus which is located in Jimma city, south western Ethiopia. This city is situated at 7°40'N and 36°50'E latitude and longitude, respectively (<http://en.wikipedia.org/wiki/Jimma>).

The *in sacco* rumen degradation study was carried out in Holeta Agricultural Research Center (HARC) animal nutrition laboratory. HARC is located in the West Shewa zone of the Oromia National Regional State, Ethiopia. It has a latitude and longitude of 9° 3' 0" N, and 38° 30' 0" E, respectively. The centre is located at an average altitude of 2391 meters above sea level (http://en.wikipedia.org/wiki/Holeta_Genet).

3.2. Description of the Browse Species

The browse species were selected based on their abundance in the area, preference and accessibility to browsing livestock as well as additional uses other than livestock feed (Yisehak *et al.*, 2010). The browse species included in the study were *Ekebergia capensis*, *Ficus sycomorus*, *Maesa lanceolata*, and *Rhus glutinosa*, which are non legume trees. These species are commonly consumed by ruminants and equines. They are available to animals throughout the year (Yisehak *et al.*, 2010).

3.3. Sampling, Drying and Storage of Plant Materials

Plant parts/morphological fractions of the tannin-rich browse species were collected from Waktola *kebele* of Omo Nada *woreda*, Jimma zone, Southwestern Oromia, Ethiopia during autumn season. Omo Nada *woreda* (http://en.wikipedia.org/wiki/Kersa_Jimma_zone) was selected based on previous study (Yisehak *et al.*, 2010). Fresh leaves and fruits were harvested from *Ekebergia capensis*, *Ficus sycomorus*, *Maesa lanceolata* and *Rhus glutinosa*, then placed in plastic bags and they were transported to JUCAVM, animal nutrition laboratory. Four individual

plants per species were sampled and analyzed individually in order to have some measure for the intra-species variation and to perform statistical analyses in factorial procedure. After arrival in 35 minutes, the plant parts were placed in a new yellow paper bags, measured (fresh weight), and placed in oven at 55°C for 72 hours (checked for a constant dry weight). The dried samples were ground to pass through 1 mm sieve size of a Wiley mill for chemical analysis and 2 mm sieve for rumen *in sacco* degradation measurement and placed in air-tight plastic sample containers and stored in the nutrition laboratory at room temperature until analysis.

3.4. Chemical Analysis

Browse species were analyzed for dry matter (DM), crude protein (CP), ether extract (EE), crude fiber (CF) and ash according to AOAC, (2005), whereas Neutral detergent fiber (NDF) was determined by the method of Van Soest *et al.* (1991), while Non-Fiber Carbohydrates (NFC) were calculated as 100-CP-NDF-ash-ether extract according to NRC, (2001). Chemical extraction of foliages for tannin components' analysis was done following the procedures of FAO/IAEA, (2002). Total condensed tannin was determined according to (Polshettiwar *et al.*, 2007). Extraction of tannins was done in 70 per cent aqueous acetone, i.e. 700 ml acetone + 300 ml distilled water. For each sample four replicates were analyzed and the absorbances of the tannin extracts were measured at 550 nm (Makkar, 1995).

3.5. In Sacco Degradability Measurements

Ruminal *in sacco* degradation of DM and OM was determined according to (Orskov and McDonald, 1979). Dried forage samples (AOAC, 2005) milled to pass through a 2 mm sieve screen was weighted (3g /bag) into 4.5 cm × 18 cm nylon bags (pore size 40 µm). The bags were manually pushed deep into the liquid phase of the ventral sac of the rumen and incubated for 6, 12, 24, 48, 72 and 96 hours in the rumen of three male Boran X Holstein-Friesian steers. Two bags were incubated for each sample in each bull for each incubation time. Upon removal from the rumen, bags were washed in running tap water while rubbing gently between thumb and fingers until the water became clear. Zero time disappearances (washing losses) were obtained by washing un-incubated bags in a similar fashion. Bags were dried in an oven at 60°C for 48 hours and weighed to determine the dry weight of the incubation residues.

In experiment the animals were allocated to a maintenance ration composed of natural pasture hay offered ad libitum as a basal ration and 2 Kg of a concentrate diet formulated from wheat bran, noug seed cake and salt in the ratio of 55: 43: 2. The animals were housed in individual pens and provided ad libitum water.

Dry matter disappearance (DMD) was estimated as follows:

$$DMD = \frac{((BW+S_1)-(BW+RW))}{(S_1 \times DM)} \times 100$$

Where, BW = bag weight; RW = residue weight; S₁ = sample weight; DM = dry matter content of the original sample

Degradability (Y) of DM was calculated using the equation of Orskov and McDonald, (1979):

$$Y = a + b (1 - e^{-ct})$$

Where “a” is the soluble fraction, “b” is the insoluble but potentially degradable fraction, “c” is the degradation rate constant of the “b” fraction, “t” is the degradation time 0, 6, 12, 24, 48, 72 and 96 hrs and the nonlinear parameters *a*, *b* and *c* were estimated using non-linear procedures of statistical analysis system (SAS, 2010).

The potential degradability was determined by the equation,

$$PD = a + b$$

The effective degradabilities (ED) for dry matter and organic matter were estimated according to ARC, (1980).

$$ED = a + bc/k + c,$$

Where k is rumen outflow rate, assumed to be 0.03/h

3.6. Statistical Analysis

A two ways analysis of variance was performed according to 4 x 2 factorial arrangements using general linear model procedure of statistical analysis system (SAS, 2010 version 9.3). Duncan's multiple range test procedure was used for mean separation. Mean differences were considered significant at $P \leq 0.05$. A spearman's correlation analysis was used to establish the relationship between some chemical composition parameters and *in sacco* ruminal DM and OM disappearances.

The DM and OM disappearances and the degradation characteristics such a , b , PD , c , ED , were estimated for each plant species and plant parts using nonlinear regression procedures of SAS.

The model used was:-

$$Y_{ij} = \mu + s_i + p_j + (sp)_{ij} + \varepsilon_{ij}$$

Where, Y_{ij} =observation, μ = population mean, S_i = i^{th} species effect ($i = 1$ to 4), $p_j = j^{\text{th}}$ plant part effect ($j = 1$ to 2), $(sp)_{ij}$ is interaction effect b/n ps and pp; and ε_{ij} is the residual error.

4. RESULTS AND DISCUSSION

4.1. The Effect of Plant Species and Forage Parts on Chemical Composition

The least square means for the chemical compositions of forage parts (leaves and fruits) of the studied plant species separately compared for the each plant species is presented in appendix Table 1. The dry matter (DM) contents varied from 92.66% in *Ficus sycomorus* leaf to 96.84% and 96.86% in *Rhus glutinosa* and *Ekebergia capensis* leaves, respectively ($P<0.001$). The crude ash (CA) content obtained in the present study for the leaf of plants fell within the range of 7.15 to 11.49 % in *Measa lanceolata* and *Ficus sycomorus* leaves, respectively. Highly significant difference was observed for CA values in the leaves of the plant species ($P<0.001$). The CP contents varied among the species and ranged from 9.76% in *Ekebergia capensis* leaf to 17.06 % in *Maesa lanceolata* leaf ($P<0.001$). The highest content of ether extract (EE) was recorded in the leaves of the three tree species such as *Maesa lanceolata* (5.17%), *Rhus glutinosa* (4.710%) and *Ekebergia capensis*(5.06%) while the lowest EE content was obtained in *Ficus sycomorus* leaf (1.57%)($P<0.001$). The neutral detergent fiber (NDF) content was ranged from 37.96% in *Ekebergia capensis* to 61.33% in *Maesa lanceolata* and 61.01% in *Ficus sycomorus* leaves, respectively ($P<0.001$). On the other hand, the highest and lowest contents of non-fiber carbohydrates (NFC) was estimated for *Ekebergia capensis* (39.30%) and *Ficus sycomorus* (13.51%) and *Maesa lanceolata* (8.71%), respectively ($P<0.001$).

Similar to chemical composition of leaves of the plants species studied, there were also significant variations in the chemical composition of fruits of the tree species for the majority of chemical composition parameters ($P<0.001$) (appendix Table 1). The chemical contents ranged from: 7.97 to 12.10% CP, 5.97 to 31% EE, 35.90 to 68.90 NDF, 5.84 to 17.28% NFE and 7.20 to 10.17% CT.

The least square mean differences for the chemical composition parameters that compared between leaves and fruits are presented in Table 1. The highest CA content was determined in the leaves of *F. sycomorus* whereas the lowest total cash values were recorded for fruits of *E. capensis* (5.43%) and *R. glutinosa*(5.57%)($P<0.001$). Similarly, the highest CP contents were

recorded in leaves as compared to fruits ($P < 0.001$). On the other hand, extremely the highest EE content was determined for fruits of *E. capensis* as compared to the rest of plant parts; EE content in the fruits tends to be higher except in the *R. glutinosa* fruit ($P < 0.001$). Similar to EE content of fruits the highest NDF content was also recorded in the fruits of the plants as compared to leaves ($P < 0.001$). Unlike the contents of EE, CF and NDF contents of the fruits, the nitrogen free carbohydrate content was found to be the highest for leaves of the *E. capensis* ($P < 0.001$). Furthermore, CT of leaves and fruits was comparable and attained the level that affects utilization of feedstuffs by livestock species. The largest ash content was obtained in the leaves of *F. sycamoros* as compared to ash values of leaves and fruits of the studied plant species ($P < 0.001$). The leaves had relatively highest CP value as compared to fruits where as the EE values in fruits tends to be larger in fruits ($P < 0.001$). On the contrary, the larger NDF content was recorded in fruits as compared leaves; the possible reason could be attributed to lack of amylase, an enzyme that catalyses the breakdown of starch into sugars enzyme digests starch during NDF analysis. Starch content decreases in the leaves during fruit development (Gerhard and Luddersl, 1997; Proietti *et al.*, 2000). The highest CT content was recorded in the leaves as compared to fruits indicating a low CT content in fruits than browse leaves.

The Crude ash (CA) content of the leaves and fruits in the present study, the mineral level in a feed and large amount of silica was approximately ranged between 5 and 12%. CA contents of browse trees ranged from 8-12% (Bogdan, 1977). Carlos *et al.*, 2005) also found 7.9 to 12.6% CA for leaves of some tree fodders.

In general, all the tree leaves and fruits had a high CP content (more than 7%) which indicates their high nutritive value in terms of CP, since the browses are intended to be used as protein supplements for low quality tropical pastures and crop by-products. Norton, (1994) reported that feeds with less than 6% CP levels are unlikely to provide the minimum ammonia levels required for maximum microbial growth in the rumen. Therefore, the entire browse species evaluated in this study would be a good protein supplements provided that they were adequately degraded and non-toxic to the rumen microbes and host animal. The preference of differences in the CP content across the plants is inconsistent with the reports of (Guerin, 1987; Mekoya *et al.*, 2008), Kind *et al.*, 2009; Ngodigha and Anyanwu, 2009) and Arigbede *et al.*, 2011). In the present

study, the CP content of the browse species was higher than the minimum level of 7–8% CP in DM required for optimum rumen function and feed intake in ruminant livestock (McDonald *et al.*, 2002; Norton, 2003; Van Soest, 1994).

Table 1. Comparison of the least square means for chemical compositions (%) of leaves and fruits of the plant species

Plant species	PP	Chemical composition, mean (%)							
		DM	CA	CP	EE	NDF	CF	NFC	CT
<i>E. capensis</i>		96.84 ^a	7.91 ^d	9.76 ^d	5.06 ^{de}	37.96 ^d	19.10 ^{bc}	39.30 ^a	8.00 ^b
<i>F. sycomorus</i>	Leaf	92.66 ^d	11.49 ^a	12.42 ^{bc}	1.57 ^f	61.01 ^b	15.40 ^{cd}	13.51 ^{bc}	8.49 ^b
<i>M. lanceolata</i>		95.10 ^b	7.15 ^e	17.06 ^a	5.17 ^{de}	61.33 ^b	21.45 ^{bc}	8.71 ^{cd}	13.65 ^a
<i>R. glutinosa</i>		96.86 ^a	9.29 ^c	16.04 ^{ab}	4.70 ^e	49.31 ^c	12.20 ^d	20.66 ^b	16.72 ^a
<i>E. capensis</i>		94.11 ^c	5.43 ^f	11.07 ^c	31.00 ^a	35.9 ^d	20.25 ^{bc}	17.28 ^b	7.27 ^b
<i>F. sycomorus</i>	Fruit	95.56 ^b	9.70 ^b	10.81 ^c	5.97 ^c	66.71 ^{ab}	27.5 ^a	9.07 ^{cd}	8.17 ^b
<i>M. lanceolata</i>		96.5 ^a	7.45 ^e	7.97 ^c	8.20 ^b	62.00 ^b	22.20 ^{ab}	14.38 ^{bc}	7.20 ^b
<i>R. glutinosa</i>		92.43 ^d	5.57 ^f	11.21 ^c	5.33 ^d	68.9 ^a	22.30 ^{ab}	5.84 ^d	8.59 ^b
	SE	0.31	0.35	0.72	1.57	2.29	1.00	1.92	0.55
	P	***	***	***	***	***	***	***	*

PP, Plant part; DM, dry matter; CA, crude ash; CP, crude protein; EE, ether extract; NDF, neutral detergent fiber; CF, crude fiber; NFC, non-fibre carbohydrates; CT, condensed tannin; ^{a, b, c, d, e, f} Least square means with the different letters within the column are significantly different ($P < 0.05$); * $P < 0.05$; *** $P < 0.001$.

The CP values obtained from the leaves and fruits in the present study were, however, lower than those CP valued reported for some leguminous browse species in south-west Nigeria (Larbi *et al.*, 1998) and also lower than the CP values obtained for *Acacia* species (Abdulrazak *et al.*, 2000). These variations could be due to plant variety, agro climatic conditions, or even maturity stages at harvesting (Makkar and Becker, 1997; Bamikole *et al.*, 2004). The chemical composition of tree leaves vary with the maturity of leaves and also with localities (Mandal, 1997). Thus, the chemical composition of the leaves of plant species analyzed here provides a good source to be used as the nutrient source of ruminant animal feed. The level of CP was, however, comparable with the reported CP of browsed species in tropical West Africa (Le- Houerou, 1980).

The entire browse species in this study had CP level within the acceptable range (7 - 14%) for production and reproduction performance of ruminants (NRC, 1981). The results of the present study also agrees with (Getachew *et al.*, 2000) in that the browse forages are better used as protein supplement than poor quality roughages such as hay, straws and stovers whose CP content is lower than 7%CP (Skerman and Riveros,1990). Voluntary feed intake rapidly falls if CP content of forage is below 6.2% (Nasrullah *et al.*, 2003). It is well reported that feeds containing less than 7% CP cannot provide the minimum ammonia levels required by rumen microorganisms to support optimum activity, the leaves and fruits of the present study can be potential supplement for low CP feeds except their CT content. Further, rumen fermentation is affected if the CP level in diet is less than 10% (Alam and Djajanigra, 1994), however, CP level in these trees is higher than this level except *M. lanceolata* fruit. Differences in CP contents between leaves of different trees are probably due to differences in protein accumulation in them during growth. Even though the CP contents of the plant parts attains the minimum protein level for optimum rumen function, their CP concentration was relatively lower than those of protein supplementary legume browses such as *Leucaena leucocephala*: 27.2%; *Sesbania sesban*: 27.6% (Mupangwa *et al.*, 2003; Thandei *et al.*, 2001; Giang *et al.*, 2004); *Albizia gummifera*: 29.6% (Yisehak *et al.*, 2010).

The NDF content of the plants, except *E.capensis* leaf (47.72%), *M.lanceolata* fruit (56.67%) and *R.glutinosa* leaf (49.50%), was found more than 60%. Optimum NDF content of a ration should be in the range of 27-30% (Jolly and Wallace, 2007). The threshold level of NDF in tropical plants beyond which feed intake of ruminants affected is 60% (Meissner *et al.*, 1991) suggesting that some of the diets included in this study marginally have above 60% NDF in DM. Tree forages with a low NDF content (20–35%) are usually of high digestibility (Norton, 1994; Bakshi and Wadhwa, 2007) that the low level of fiber can facilitate the colonization of the plant parts included in the present study by the rumen microbial population, which in turn might induce even higher fermentation rate, therefore improving its digestibility(Van Soest, 1994). The results of present studies were inconsistent with (Schmidek *et al.*, 2000) and (Cheema *et al.*, 2011) who reported 26.2 to 39.3% NDF for multipurpose trees and shrub species. The NDF values of the plants of the present study was also comparable with the NDF values reported for

some browse species in Nigeria (Larbi *et al.*, 1998) and shrub species from a mountain area in northern Spain (Frutos *et al.*, 2000) with just little variations. High contents of cell wall are typical of tropical forages (Van Soest, 1982) which have serious implication on the digestibility of forages. NDF actually determines the rate of digestion because it is inversely related to digestibility (McDonald *et al.*, 1995; Gillespie, 1998).

The EE content in leaves of the plants except in *F.sycomorus* (avg. 1.57%) exceeded 5%. The EE content (avg. < 5% in DM) is an indication of low energy level for the animal. Odedire and Babayemi (2008) reported that feedstuffs contained more than 5% considered higher energy level for an animal. Therefore the present study could achieve the above requirement. However, the EE content of *E. capensis* fruits was above the acceptable range for the animals (avg. 28.10%). Total diets that do not contain EE more than 10 % (Preston, 1995). The higher value of ether extracts in some of the tested samples is an indication of higher energy level for the animal (Babayemi and Bamikole, 2006; Odedire and Babayemi, 2008) and this is a major form of energy storage in plants which is being utilized by the animals for body maintenance and production. The crude fat content of all the tree fruits and leaves evaluated were higher than the levels reported in pasture grass (0.6-1.3 per cent), and fodder maize (4.4 per cent) (Schlink and Burt, 1993).

The CT content in all the plants was above 5% that is why these plants considered as tannin rich plants. Tannin concentrations higher than 5% adversely affect forage intake and digestibility (Perevolotsky *et al.*, 1993; Silanikove *et al.*, 1996). Tannins have ability to bind and inhibit the digestive enzyme activities (Kumar and Singh, 1984) and affect the microbial and enzyme activities (Makkar *et al.*, 1989), whereas, lower concentration of tannins can improve nutrition for ruminants by reducing protein degradation in the rumen and increasing the flow of amino acids to the intestine (Mc Nabb *et al.*, 1996). Concentration of tannins less than 4% in the ration is beneficial by promoting bypass protein and bloat suppression in ruminant animals (Aganga and Tshwenyane, 2003). Compared with tropical, mature grasses, browse appears to be richer in protein and minerals (Le Houerou, 1980; Devendra, 1995). In most situations, its practical use is as a supplement to enhance the intake and utilization of other fibrous crop residues like cereal straws and hays, and thus meet the maintenance and variable levels of production requirements.

4.2. The Effect of Plant Species and Plant Parts on *In Sacco* Ruminal Dry Matter Disappearances

Interspecies variation for DM disappearances from leaves and fruits of tannin rich trees for different incubation hours are presented in appendix Table 2. The highest DM disappearance at zero hour incubation was recorded in leaves of *E.capensis* (29.43%) whereas the DM disappearance value was lowest for *F.sycomorus* leaves (6.32%) ($P<0.001$). This might be attributed to processing or distribution in particle size of the ground materials. The reports of Welch, (1986); Singh *et al.*, (1989) and Khan *et al.*, (2009) confirmed that DM disappearance at zero hour is mainly due to mechanical action rather than microbial fermentation. At 6 hours of post incubation, the highest DM disappearances were recorded for the leaves of *E. capensis* (32.12%) and *M. lanceolata* (32.60%) where as the lowest DM disappearances were determined for *F. sycomorus*(15.82%) and *R. glutinosa* (16.87%) leaves ($P<0.001$). Yet at 6 hour incubation, the *E. capensis* fruits had the highest DM disappearances as compared to rest of the plants ($P<0.001$). Across all the incubation hours, the leaves of *E. capensis* had the highest DM disappearances; comparably, *M. lanceolata* leaves had also the highest DM disappearances in 6, 24 and 72 hours incubation in contrast to the rest of plants ($P<0.001$). In comparison, the highest DM disappearance value was only recorded for *E. capensis* fruit ($P<0.001$).

The least square mean differences for the DM disappearance that compared between leaves and fruits are presented in Table 2. At zero-hour incubation both the highest and lowest DM disappearance or bag wash loss was recorded in *E.capensis* fruit (39.28%) and leaves of *F.sycomorus* (6.32%), respectively ($P<0.001$). *E.capensis* leaves and fruits acquire the largest share in DM disappearances in almost all incubations hours as compared to DM disappearances in edible parts of the rest plants ($P<0.001$). In general species variation was significant for DM disappearances across the incubation hours. A possible reason to this effect might be species difference in since plant and their edible fractions has inconsistent values of chemical constituents used for rumen microbes. Arigbede *et al.* (2011) reported similar findings that an effect of species variation is significant for variations in DM disappearances of various multipurpose tree species.

Table 2. Least square means for *in sacco* ruminal dry matter disappearances compared for leaves and fruits of the plant species

Plant species	PP	Incubation time (h), mean (%)						
		0	6	12	24	48	72	96
<i>E. capensis</i>		29.43 ^b	32.12 ^b	59.47 ^a	73.11 ^a	82.49 ^a	81.79 ^a	83.38 ^a
<i>F. sycomorus</i>	Leaf	6.32 ^g	15.82 ^d	26.09 ^c	22.53 ^d	61.07 ^b	59.52 ^b	69.89 ^c
<i>M. lanceolata</i>		23.69 ^c	32.60 ^b	35.59 ^b	64.99 ^{ab}	65.72 ^b	81.46 ^a	77.53 ^b
<i>R. glutinosa</i>		19.28 ^{de}	16.87 ^d	17.99 ^d	23.72 ^d	38.20 ^c	47.07 ^c	47.64 ^d
<i>E. capensis</i>		39.28 ^a	43.34 ^a	60.01 ^a	62.02 ^b	78.95 ^a	75.94 ^a	84.85 ^a
<i>F. sycomorus</i>		20.81 ^d	23.29 ^c	34.20 ^b	43.17 ^c	58.90 ^b	62.34 ^b	69.89 ^c
<i>M. lanceolata</i>	Fruit	10.32 ^f	15.14 ^d	19.03 ^d	28.64 ^d	40.11 ^c	54.15 ^{bc}	46.08 ^d
<i>R. glutinosa</i>		17.60 ^e	17.91 ^{bcd}	20.30 ^{cd}	22.16 ^d	26.46 ^d	34.16 ^d	21.65 ^e
	SE	2.04	1.55	2.47	3.14	2.99	2.51	3.06
	P	***	***	***	***	***	***	***

PP, plant parts; ^{a, b, c, d, e, f, g} Least square means in a column with different superscripts are significantly different ($P < 0.05$); *** $P < 0.001$; SE, Standard error of means

The other possible reason might be associated with the NFC content, the major energy source for rumen microbes, in content in the leaves and fruits of *E. capensis* were found to be superior. The range of NFC content showed that the trees under evaluation can be easily degraded or fermented because NFC is an estimate of the carbohydrate pool that differ in digestibility from CF and NDF. It has also been reported that NFC has a positive relationship with ammonia nitrogen (NH₃-N) utilization in the rumen (Tylutki *et al.*, 2008). As nitrogen utilization by rumen microflora is related to the amount of fermentable energy, the adequate NFC contents especially in *E. capensis* could enable efficient microbial protein synthesis by promoting better utilization of rumen ammonia released from feeds with high content of rumen degradable CP (Cabrita *et al.*, 2006). DM disappearance values were highest at terminal incubation hours across plant species. Many authors (Orskov *et al.*, 1988; Kabuga and Darko, 1993; Tesema and Baars, 2003; Lanyasunya *et al.*, 2006; Vranic *et al.*, 2009 and Lebopa *et al.*, 2011) confirmed the increasing trends of that ruminal degradation of DM as advances in incubation hours. The other possible reason could be tannin content of the feed materials. Although the tannin content of the leaves and fruits are attained the level that affects nutrient utilization, CT content in leaves and fruits of *E. capensis* was one of the lowest CT values among the plant parts. Tannins are complex polyphenolic compounds with an ability to precipitate proteins and to form complexes with

carbohydrates thereby, reducing the digestibility (Kumar and Vaithiyanathan, 1990; Makkar, 2003; Ferreira, 2004; Mezzomo *et al.*, 2011).

Except in fruits and leaves of *E. capensis* and leaves of *M. lanceolata*, the DM disappearance of the plants is lower than the values recorded for most of the legume and non-legume multipurpose trees reported by Ramana *et al.* (2000). This difference might be associated to differences in plant species. The higher NDF and CT contents in the plant species might also attribute to lower DM disappearances.

4.3. In Sacco Dry Matter Degradability Characteristics of the Leaves and Fruits of the Plant Species

The *in sacco* DM degradability characteristics of leaves and fruits of the studied plants are presented in appendix Table 3. The highest and lowest soluble fraction (b) of leaves was recorded in *M. lanceolata* (14.55%) and *E. capensis* (-16.20%), respectively (P<0.001). This could be related to the loss of finer particles from the bags in this treatment, rather than a higher solubility. Yet in the fruit, the highest value of soluble fraction (35.51%) was obtained for *E. capensis*. Except in fruits of *M. lanceolata* (14.55%) and *F. sycomorus* (15.49%), the values of soluble fraction (a) in the present study were lower than the 'a' values estimated for 20 multipurpose trees and shrub species (both legumes and non legumes) studied in Nigeria by Ngodigha and Anyanwu, (2009). The possible reason could be linked to variation in plant species. However, the least square means obtained for slowly degradable fraction (b) in the leaves of *E. capensis* was found to be the superior (98.37%) where as the least value was recorded for *R. glutinosa* leaves (P<0.001). This could be associated with the concentration of CTs; the highest CT concentration in *R. glutinosa* leaves has had the least DM disappearance of the potentially degradable fraction. CTs depress DM degradability (Gonzalez *et al.*, 2002; Ozkan and Sahin, 2006; Gupta *et al.*, 2011). In general, *E. capensis* leaves had the highest values for slowly degradable fraction, effective degradability and rate of degradation (c) as compared to values recorded for other plants (P<0.001). Unlike in leaves, *E. capensis* fruit had significantly the highest a soluble fraction (a), potential degradability (b) and effective degradability (ED) values as compared to the 'a', PD and ED values in the fruits of other plants (P<0.001). This might be due higher content NFC and as well as EE content in *E. capensis* leaves and fruits. Higher level

of soluble fraction is known to result in a more efficient fermentation in the rumen (Beever *et al.*, 1978). The differences in soluble fraction could be attributed to the proportion of soluble carbohydrates to structural carbohydrates (Ngodigha and Oji, 2009). According to Van Soest (1982), the soluble carbohydrates ferment faster than structural carbohydrates.

Table 3. Comparison of the least square means for in sacco ruminal drymatter degradability characteristics of leaves and fruits of the plant species

Plant species	PP	Degradability parameters				
		a	b	PD	ED	c
<i>E. capensis</i>		-16.2 ^h	98.37 ^a	82.17 ^c	61.83 ^b	0.12 ^a
<i>F. sycomorus</i>		7.37 ^f	77.82 ^b	85.19 ^a	35.78 ^e	0.02 ^e
<i>M. lanceolata</i>	Leaf	14.5 ^c	64.91 ^c	79.47 ^d	53.76 ^c	0.05 ^b
<i>R. glutinosa</i>		11.23 ^e	52.76 ^e	63.99 ^f	27.86 ^g	0.01 ^f
<i>E. capensis</i>		35.51 ^a	47.11 ^f	82.62 ^b	63.18 ^a	0.04 ^c
<i>F. sycomorus</i>		15.49 ^b	56.15 ^d	71.64 ^e	43.28 ^d	0.03 ^d
<i>M. lanceolata</i>	Fruit	5.78 ^g	46.59 ^g	52.36 ^g	29.22 ^f	0.03 ^d
<i>R. glutinosa</i>		13.92 ^d	13.69 ^h	27.61 ^h	22.57 ^h	0.05 ^b
	SE	2.63	4.69	3.99	3.11	0.006
	P	***	***	***	***	***

PP, plant part; a, soluble fraction; b, slowly degradable fraction; (%/hour); PD, potential degradability; ED, effective degradability; c, rate of degradation; ^{a, b, c, d, e, f, g, h} Least square means within the same column with different superscripts are significantly different ($P < 0.05$); SE, Standard error of the mean; *** $P < 0.001$

The *in sacco* DM degradability characteristics of fruits and leaves of various multipurpose tree species (MPTS) compared and the results were presented in Table 3. Although the species variation was significant for degradability parameters, the effect of plant parts for the same species was also found to be significant for the degradability parameters ($P < 0.001$). The difference among the MPTS in degradation characteristics may be partly related to variations in chemical composition (Nashlai *et al.*, 1994), the relationship of cell wall polysaccharides and their effect on rumen microbial attachment and colonization of digesta particles (Cheng *et al.*, 1984). The high ED recorded for *E. capensis* may relate to its low cell contents (NDF). This confirmed earlier reports that *in situ* nylon bag method could be used to rank MPTS for quality in initial screening studies. Based on ED values in this study, MPTS could be ranked as high ($> 450 \text{ g kg}^{-1} \text{ DM}$), medium ($400\text{-}450 \text{ g kg}^{-1} \text{ DM}$) and low ($< 400 \text{ g kg}^{-1} \text{ DM}$) quality groups. *E. capensis* (leaf and fruit), *M. lanceolata* (fruit), were among the high quality group; *F. sycomorus*

fruit was in the medium quality group, while both *R. glutinosa* leaf and fruit and *M.lanceolata* fruit belonged to the low quality group. The highest (0.12%/h) and lowest (0.01%/h) values of passage rate were recorded for *E.capensis* leaves and *R.glutinosa* leaves as compared to the 'c' values in fruits of the plant species, respectively (P<0.001).

4.4. The Effect of Plant Species and plant parts on *in sacco* ruminal Organic Matter Disappearances

The 0- hour disappearance of OM was largest for leaves and fruits of *E.capensis* as compared to the rest of plants (P<0.001) (appendix Table 4). Similarly, at 6 h incubation the highest OM disappearance was recorded for leaves and fruits of *E.capensis*, as well as leaves *M.lanceolata*. Throughout the incubation hours the leaves and fruits of *E.capensis* has taken the highest OM disappearance values followed by leaves of *M.lanceolata* as compared to OM disappearance values of plant parts in the rest of plant species(P<0.001).

The high washing loss (0 h disappearance) of MPTS evaluated in this study is an indication of the presence of soluble or ruminally degradable nutrients that may be rapidly utilized in the rumen. The MPTS recorded more than 60% OM degradability at 24 h which implied that they were extensively degraded in the rumen. As compared to 0h OM disappearances of leaves and fruits, the highest value (39.28%) was recorded in the *E.capensis* fruit (P<0.001, Table 4). Yet, *in sacco*, throughout the incubation hours other than 0 h, even though the OM disappearance values were one of the highest values among plants and their edible fractions, statistically significant variation was not recorded between leaves and fruits of *E.capensis* (P>0.001). Variation in protein degradability is directly related to the proportion of structural and non-structural protein and carbohydrate fractions which in turn affects their solubility and bioavailability (Whetton *et al.*, 1997). The lowest OM degradability values for leaves and fruits (< 60%) of the plant species except *E.capensis* leaves and fruits, as well as *M.lanceolata* leaves might be associated to the presence of secondary metabolites. Several studies have documented effects of secondary metabolites especially tannins (Reed, 1995; Ammar *et al.*, 2004; Gasmi-Boubaker *et al.*, 2005) on OM degradability; However, it can be concluded that tannins at levels reported in this study could not affect OM degradability in *E.capensis* leaves and fruits and leaves of *M.lanceolata*, even if CT content reached the values that complicates feed digestion.

Although, changes in the tannin content of individual species may affect rumen nitrogen and OM degradability (Barry and Forss, 1983), it is perhaps not valid to compare the effect of tannin content on such parameters between species as there is no existing accurate method for estimation of "active" tannins. The other possible reason might be associated with higher NDF contents; ruminant livestock require fiber for normal rumen function but fiber also limits feed intake and digestibility (Albrecht and Broderick, 1990).

Table 4. Comparison of least square means of leaves and fruits of the tannin rich plants for *in sacco* organic matter disappearances

Plant species	PP	Incubation time (h), mean (%)						
		0	6	12	24	48	72	96
<i>E. capensis</i>		29.43 ^b	37.55 ^{ab}	61.05 ^a	73.28 ^a	82.48 ^a	81.79 ^a	83.38 ^a
<i>F. sycomorus</i>	Leaf	6.32 ^g	22.63 ^{cd}	26.45 ^c	23.57 ^c	62.64 ^c	61.05 ^b	69.89 ^b
<i>M. lanceolata</i>		23.69 ^c	31.21 ^{bc}	36.05 ^b	65.91 ^a	72.30 ^b	81.69 ^a	77.60 ^a
<i>R. glutinosa</i>		19.28 ^{de}	17.44 ^d	20.63 ^c	26.26 ^c	42.02 ^e	47.27 ^d	48.98 ^c
<i>E. capensis</i>		39.28 ^a	43.34 ^a	60.68 ^a	64.81 ^a	81.34 ^a	80.29 ^a	84.99 ^a
<i>F. sycomorus</i>	Fruit	20.81 ^d	21.27 ^d	34.96 ^b	45.43 ^b	53.40 ^d	64.88 ^b	69.36 ^b
<i>M. lanceolata</i>		10.32 ^f	14.85 ^b	19.03 ^c	29.73 ^c	47.86 ^{de}	54.61 ^c	45.63 ^c
<i>R. glutinosa</i>		17.60 ^e	17.91 ^d	20.30 ^c	24.39 ^c	27.73 ^f	38.25 ^e	31.09 ^d
	SE	2.04	1.73	2.49	3.06	2.83	2.37	2.82
	P	***	***	***	***	***	***	***

PP, plant part; ^{a, b, c, d, e, f, g} Least square means within the same column with different superscripts are significantly different ($P < 0.05$); *** $P < 0.001$; SE, Standard error of the mean

4.5. Effect of Species and Plant Parts on *In Sacco* Organic Matter Degradability

Table 5 shows the nonlinear parameter estimates and effective degradability values of OM of the leaves and fruits of the studied plants. The soluble fraction (a) ranged from -1.02% in *R. glutinosa* to 10.69% in *E. capensis* leaves where the variation was significant for the plant species ($P < 0.001$). On the other hand, the highest and lowest 'a' value was recorded for fruits of *R. glutinosa* (31.84%) and *E. capensis* (14.22%) ($P < 0.001$). The insoluble but degradable fraction (b) was 83.35% in *R. glutinosa* and 45.21% in *E. capensis* leaves ($P < 0.001$); On the contrary, the highest and lowest values of potential degradability (PD) was recorded for *F. sycomorus* (89.89%) and 55.90% for *E. capensis* ($P < 0.001$) respectively. Similarly, the highest and lowest effective degradability (ED) was obtained in *R. glutinosa* (63.97%) and *E. capensis* leaves

($P < 0.001$). The degradation rate constants (c) varied widely between MPTS with similar rates for *E.capensis* and *F.sycomorus*. The highest washing losses, soluble or rapidly degradable OM fraction at time 0h was highest for *R.glutinosa* fruit whereas it was the least value (-1.02%) in the leaf of the same plant ($P < 0.001$) (Table 5). This might be associated to differences in the solubility of leaves and fruits of the same plant. Yet, in leaves and fruits, the value recorded for the insoluble but potentially fermentable OM fraction that degrades with time was highest in *R.glutinosa* (83.35%) leaves and lowest in *E.capensis* fruit (45.21%) ($P < 0.001$). Conversely, among the plant species, the highest PD (89.89%) and lowest (35.60%) was determined for *F.sycomorus* leaves and *E.capensis* fruit ($P < 0.001$). Similar to the rapidly soluble fraction 'a', effective degradability (ED) was found to be largest in fruits as compared to leaves of the plants ($P < 0.001$). In general, it was observed that variation of plant parts lead to highly significant differences in OM degradability characteristics of the plant species.

The observed differences in OM degradation characteristics of the studied plant species may be partly due to the differences in their chemical composition which is influenced by the species, foliage parts of the trees. The range of values for the readily soluble 'a', potentially degradable fractions 'b', the rate 'c' and extent of OM degradation in this study agreed with the reports of (Larbi *et al.*, 1998; El Hassan *et al.*, 2000; Hervas *et al.*, 2000) which also confirmed significant differences in OM degradation characteristics of several tropical browse trees. Degradation constants as measured by the *in sacco* nylon bag technique are strongly related to digestible DM intake (Kibon and Orskov, 1993). Thus, interspecies variations in OM degradability could result in differential intakes of the plants when given as sole diets to animals. The 'a' for leaves of the plants are lower than previous reports for various MPTS in Nigeria (Ngodigha and Anyanwu, 2009; Arigbede *et al.*, 2011), 12 species in Kenya (Ondiek *et al.*, 2010). These differences might be attributed to differences in plant species location and contents of secondary plant components especially condensed tannin; higher CT content is obtained in the present study.

Table 5. Comparison of least square means for *in sacco* ruminal degradability characteristics of organic matter of leaves and fruits of the plant specie

Plant species	PP	Degradability parameters				
		a	b	PD	ED	C
<i>E. capenesis</i>		10.69 ^e	45.21 ^g	55.90 ^f	29.61 ^g	0.02 ^e
<i>F. sycomorus</i>		12.63 ^d	77.27 ^b	89.89 ^a	38.11 ^e	0.02 ^e
<i>M. lanceolata</i>	Leaf	8.11 ^f	71.98 ^c	80.09 ^d	54.55 ^c	0.05 ^b
<i>R. glutinosa</i>		-1.02 ^h	83.35 ^a	82.33 ^c	63.97 ^b	0.11 ^a
<i>E. capenesis</i>		14.22 ^c	21.38 ^h	35.60 ^h	24.58 ^h	0.03 ^d
<i>F. sycomorus</i>	Fruit	15.26 ^b	56.40 ^d	71.66 ^e	42.84 ^d	0.03 ^d
<i>M. lanceolata</i>		2.06 ^g	50.28 ^f	52.34 ^g	30.59 ^f	0.04 ^c
<i>R. glutinosa</i>		31.84 ^a	52.02 ^e	83.86 ^b	64.65 ^a	0.05 ^b
	SE	2.03	4.04	3.82	3.15	0.006
	P	***	***	***	***	***

PP, plant part; 'a', soluble fraction; b, slowly degradable fraction; PD, potential degradability; ED, effective degradability; c, rate of degradation; ^{a, b, c, d, e, f, g, h} Means within the same column with different superscripts are significantly different ($P < 0.05$); *** $P < 0.001$; SE, standard error of the mean.

4.6. The Correlation between Chemical Composition and *In Sacco* Ruminal Degradation of Dry matter and Organic Matter

The spearman's correlation coefficient (r) between chemical composition and *in sacco* ruminal degradability of DM and OM are presented in Table 6. The statistical correlation between CT and DM and OM disappearances in all incubation hours was negative and highly significant ($P < 0.001$) (Table 6). This is an indication of the negative influences of CTs on DM and OM disappearances the MPTS. Condensed tannins are known to reduce digestible matter by binding essential nutrients in the digestive tracts of livestock (Hagerman and Butler, 1981; Provenza *et al.*, 1990; Allredge, 1994; Reed, 1995; Getachew *et al.*, 2000; Makkar, 2003). The poor non-significant correlation was observed between CP and *in sacco* degradability of DM and OM, might be associated with the condensed tannin content in the MPTS that in turn affects degradability by protecting feed proteins from microbial utilization. Moreover, the correlation between DM and OM disappearances and NDF or CF was negative; however, the strong and significant correlation was observed for DMD at ($r = 0.755$) and OMD ($r = 0.786$) at 96 h ($P < 0.05$). The digestibility of plant material in the rumen is related to the proportion and lignification of cell walls (Van Soest, 1994). NDF actually determines the rate of digestion because it is inversely related to digestibility (McDonald *et al.*, 2002). Except in leaves and fruits

of *E. capensis* and *R. glutinosa* leaves the NDF contents of the plant species are more 60 % in DM and degradability was associated with higher NDF contents. Ruminant livestock require fibre for normal rumen function but fibre also limits feed intake and digestibility (Albrecht and Broderick, 1990).

6. The spearman's rho correlation coefficient between chemical composition and *in sacco* dry matter and organic matter disappearances for some incubation hours

	DMD ₁₂	DMD ₂₄	DMD ₄₈	DMD ₇₂	DMD ₉₆	OMD ₁₂	OMD ₂₄	OMD ₄₈	OMD ₇₂	OMD ₉₆	DM	Ash	CP	EE	NDF	CF	NFC	CT	
DMD ₁₂	1.00	0.74*	0.89**	0.86**	0.90**	0.91**	0.71*	0.88**	0.86**	0.88**	-0.21	-0.33	0.12	0.29	-0.48	0.07	0.12	-0.36**	
DMD ₂₄		1.00	0.83*	0.93**	0.80*	0.81*	0.98**	0.83*	0.93**	0.76*	0.43	-0.19	0.21	0.21	-0.50	-0.02	0.41	-0.31**	
DMD ₄₈			1.00	0.95**	0.95**	0.91**	0.74*	1.00**	0.95**	0.95**	0.10	-0.07	0.14	0.02	-0.69	-0.26	0.43	-0.38**	
DMD ₇₂				1.00	0.90**	0.90**	0.86**	0.95**	1.00**	0.88**	0.19	-0.05	0.12	0.05	-0.52	-0.10	0.31	-0.29**	
DMD ₉₆					1.00	0.95**	0.79*	0.95**	0.90**	0.99**	0.08	-0.13	0.01	0.08	-0.76*	-0.29	0.42	-0.25**	
OMD ₁₂						1.00	0.76*	0.91**	0.91**	0.93**	0.10	-0.10	0.02	-0.02	-0.64	-0.21	0.36	-0.12**	
OMD ₂₄							1.00	0.74*	0.86**	0.67	0.41	-0.33	-0.24	0.31	-0.41	0.10	0.33	-0.29**	
OMD ₄₈								1.00	0.95**	0.95**	0.10	-0.07	0.14	0.02	-0.69	-0.26	0.43	-0.38**	
OMD ₇₂									1.00	0.88**	0.19	-0.05	0.12	0.05	-0.52	-0.10	0.31	-0.29**	
OMD ₉₆										1.00	0.05	-0.12	0.05	0.024	-0.79*	-0.36	0.43	-0.24	
DM											1.00	0.31	0.24	-0.14	-0.33	-0.33	0.71*	-0.02	
Ash												1.00	0.02	-0.64	0.07	-0.29	0.17	0.19	
CP													1.000	-0.52	-0.05	-0.38	-0.33	0.88**	
EE														1.00	0.14	0.67	-0.14	-0.64	
NDF															1.00	0.76*	-0.81*	0.17	
CF																1.00	-0.67	-0.29	
NFC																	1.00	-0.31	
CT																			1.00

DMD₁₂, dry matter disappearance at 12 h; *DMD₂₄*, dry matter disappearance at 24 h; *DMD₄₈*, dry matter disappearance at 48 h; *DMD₇₂*, dry matter disappearance at 72 h; *DMD₉₆*, dry matter disappearance at 96 h; *OMD₁₂*, organic matter disappearance at 12 h; *OMD₂₄*, organic matter disappearance at 24 h; *OMD₄₈*, organic matter disappearance at 48 h; *OMD₇₂*, organic matter disappearance at 72 h; *OMD₉₆*, organic matter disappearance at 96 h; *DM*, dry matter; *CA*, crude ash; *CP*, crude protein; *EE*, ether extract; *NDF*, neutral detergent fiber; *CF*, crude fiber; *NFC*, non-fibre carbohydrates; *CT*, condensed tannin; **P*<0.05; ***P*<0.0

5. SUMMARY AND CONCLUSION

The present study covered chemical composition, *in sacco* rumen dry matter and organic matter degradability of leaves and fruits of indigenous tannin rich multipurpose tree species (MPTS) and also assessed the relationship between chemical composition and *in sacco* ruminal degradability parameters

The chemical composition of the multipurpose tree species has wide-ranging variation both in plant species and edible plant parts. The leaves of the plants were rich in crude ash, crude protein, non-fiber carbohydrates and condensed tannin when comparing with the fruits. In contrast, fruits had the highest ether extract, neutral detergent fiber and crude fiber comparing with the leaves of the plants.

The present study shows that all the MPTs are potential sources for use as protein banks to supplement grasses or crop residues in dry seasons. However, the nutrient activating factor (condensed tannin) in the edible parts of studied plants were greater than the threshold value that affects nutrient utilization and bioavailability in grazing livestock species. yet, the presence of CT at higher levels in some species may be a major constraint to their utilization by the livestock.

Dry matter and OM degradation were highly correlated with time taken for the incubation. *In sacco* degradability trial showed presence of both ruminally degradable and undegradable dietary nutrients. Rumen degradation characteristics could be used to characterize and detect variation in forage quality of the browse. Depending on their DM disappearance potentials both leaf and fruit of *E. capensis* were $> M. Lanceolata > F. sycomorus > R. glutinosa$ at 96hr of incubation. While for OM disappearance leaves of *E. capensis* and $M. lanceolata > F. sycomorus > R. glutinosa$ leaves but for the fruits, $E. capensis > F. sycomorus > M. lanceolata > R. glutinosa$. So it is possible to conclude leaves and fruits of *E. capensis* are superior in DM and OM disappearances of the other plant species. The lowest DM and OM were resulted in both leaf and fruit of *R. glutinosa* plant at the end of 96h incubation time. In general, the DM and OM disappearances of plant parts were negatively correlated to concentrations of condensed tannins.

Among the plant species, the highest PD (89.89%) and lowest (35.60%) was determined for *F. sycamoros* leaves and *E.capensis* fruit ($P<0.001$). Similar to the rapidly soluble fraction 'a', effective degradability (ED) was found to be largest in fruits as compared to leaves of the plants ($P<0.001$). In general, it was observed that variation of plant parts lead to highly significant differences in OM degradability characteristics of the plant species.

Based on the above summary and conclusion the following recommendations are forwarded:

- The chemical composition can indicate nutrient potentials of a given feedstuff but depending on the simple chemical composition parameters like in the present study it is difficult to recommend a given plant is better than the other, thus the concise recommendation should be based on the complete chemical, allelochemical and/or phytochemical parameters before including this plant species in to total mixed rations are necessary.
- Since the present works only reveal the chemical composition and *in sacco* degradation of plant parts in one season that can again not clearly show the seasonal influences on the chemical composition and *in sacco* degradation of edible fractions of plant species; thus combining the effects of seasons and plant parts should call for further study.
- To see the clear effect of CTs on feed degradability, the CP and NDF degradability should be done because tannin bind both protein and carbohydrates.
- Further studies should again focus on the bioavailability of nutrients using CT rich MPTS in different classes and breeds of livestock species.

6. REFERENCES

- Abdulrazak, S. A., Fajihara, T., Ondiek, J.K. and Orskov, E.R., 2000. Nutritive evaluation of some Acacia tree leaves from Kenya. *Anim. Feed Sci. Technol.* **85**: 89-98.
- Abiliza, E., Kimambo, A.E. and Muya, H. M. H., 1991. Rumen degradation of dry matter and organic matter of different parts of the banana plant. Sokoine University of Agriculture, Morogoro, Tanzania.
- Aganga, A.A. and Tshwenyane, S.O., 2003. Feeding value and anti-nutritive factors of forage tree legumes. *Pak. J. Nutr.* **2**: 170-177.
- Agricultural and Food Research Council., 1993. Agricultural and food research council: technical committee on responses to nutrients. Nutrients requirements of ruminant animals: protein. *Nutr Abst Rev.* **9(2)**: 65-71.
- Ahn, J.H., Robertson, B.M., Elliot, R., Gutteridge, C. and Ford, C.W., 1989. Quality assessment of tropical browse legumes: tannin content and protein degradation. *Animal Feed Science and Technology.* **27**: 147-156.
- Alam, M.P. and Djajanigara, A., 1994. Nutritive value and yield of potential tree leaves and shrubs in Bangladesh. In: *Proceedings of 7th AAAP Animal Science Congress on Sustainable Animal Production and Environment*. PP. 317-318.
- Albrecht, K.A. and Broderick, G.A., 1990. Degradation of forage legume protein by rumen microorganism. In: *Agronomy Abstract*. American Society of Agronomy, Madison, WI. p.185.
- Alemayehu, M., 1998. Natural Pasture Improvement Study around Smallholder Dairy Areas. MoA Small Dairy Development Project (SDDP), Addis Ababa, Ethiopia.
- Alemayehu, M., 2002. Forage Production in Ethiopia: A case study with implications for livestock production. Ethiopian Society of Animal Production (ESAP), Addis Ababa, Ethiopia.
- Alemayehu, M., 2003. Country pasture/Forage resources profiles: Ethiopia. Food and Agriculture Organization of the United Nations (FAO).
Web site: <http://www.fao.org/ag/AGP/AGPC/doc/Counprof/Ethiopia/Ethiopia.htm>
- Allredge, J., 1994. Effects of condensed tannins on browsers and grazers; qualitative and quantitative defense. Colorado State University, Fortcollins, Colorado.
- Alonso-Diaz, M.A., Torres-Acosta, J.F.J., Sandoval- Castro, C.A. and Hoste, H., 2010. Tannins in tropical tree fodders fed to small ruminants: A friendly foe? *Small Rumin Res.* **89(2-3)**:164-173.

Ammar, H., Lopez, S., Gonzalez, J.S. and Ranilla, M.J., 2004. Comparison between analytical methods and biological assays for the assessment of tannin related anti-nutritive effects in some Spanish browse species. In: *J. Sci. Food Agric.* **84**: 1349-1356.

Anele, U. Y.; Arigbede, O. M.; Olanite, J. A.; Adekunle, I. O.; Jolaosho, A. O.; Onifade, O. S. and Oni, A. O., 2008. Early growth and seasonal chemical composition of three indigenous multipurpose tree species (MPT) *Moringa oleifera*, *Millettia griffoniana* and *Pterocarpus santalinoides* in Abeokuta, Nigeria. *Agroforestry Systems.* **73**: 89-98.

AOAC (Association of Official Analytical Chemists), 1990. Official Methods of Analysis of the Association of Official Analytical Chemists, 14th ed. Association of Official Analytical Chemists, Washington, DC.

AOAC (Association of Official Analytical Chemists), 1990. Official Methods of Analysis of the Association of Official Analytical Chemists, 15th ed. Association of Official Analytical Chemists, Washington, DC.

AOAC (Association of Official Analytical Chemists), 1997. Official Methods of Analysis. 16th Ed. Association of Official Analytical Chemists, Arlington, VA, USA.

AOAC (Association of Official Analytical Chemists), 2005. Official Methods of Analysis of the Association of Analytical Chemists, 18th Ed. Association of Official Analytical Chemists, Gathersburg, MD, USA.

ARC (Agricultural Research Council), 1980. The nutrient requirement of ruminant livestock. Technical review by Agricultural Research Council Working Party. Commonwealth Agriculture Bureau, Farnham Royal, UK.

ARC (Agricultural Research Council), 1984. The Nutrient Requirements of Ruminant Livestock. Supplement 1. Slough, UK: Commonwealth Agricultural Bureau.

Aremu, O.T. and Onadeko, S.A., 2008. Nutritional ecology of African buffalo (*Syncerus caffer nanus*). *Int. J. Agric. Res.* **3**: 281-286.

Arigbede, O.M., Anele, U. Y., Sudekum, K.H., Hummel, J., Oni, A. O., Olanite, J. A., Isah, A. O., 2011. Effects of species and season on chemical composition and ruminal crude protein and organic matter degradability of some multi-purpose tree species by West African dwarf rams. *Journal of Animal Physiology and Animal Nutrition.* DOI: 10.1111/j.1439-0396.2011.01146.x.

Armenta, J.A., Quintana, R.G. and Ramirez, R., 2009. Organic Matter Degradability of Diets by Range Goats. *Journal of Animal and Veterinary Advances.* **8(5)**: 825-828.

Asiegbu, J. E. and Anugwa, F. O. I., 1988: Seasonal availability, physical and chemical characteristics of four major browse plants used for stall-feeding of livestock in Eastern Nigeria. *Bulletin of Animal Health and Production in Africa.* **36**: 221–230.

Azene Bekele-Tesemma., 1993. Useful trees and shrubs for Ethiopia: Identification, propagation and management for agricultural and pastoral communities. SIDA.Nairobi, Kenya.

Babayemi, O.J and Bamikole, M.A., 2006. Supplementary value of *Tephrosia bracteolata*, *Tephrosia candida*, *Leucaena leucocephala* and *Gliricidia sepium* hay for West African dwarf goats kept on range. *Journal of Central European Agriculture*. **7 (2)**: 323-328.

Babayemi, O.J., Demeyer, D. and Fievez, V., 2004b. *In vitro* fermentation of tropical browse seeds in relation to their content of secondary metabolites, *Journal of Animal and Feed Science*. **13(1)**: 31-34.

Bakshi, M.P.S. and Wadhwa, M., 2007. Tree leaves as complete feed for goat kids. *Small Rumin.Ref* **69**: 74-78.

Bamikole, M.A., Ikhatua, O.M., Arigbede, O.M., Babayemi, O.J. and Etela, I., 2004. An Evaluation of the Acceptability as forage of some Nutritive and Antinutritive Components and of the Dry Matter Degradation Profiles of Five Species of Ficus. *Trop. Anim. Health Prod.* **36**: 157-167.

Barry, T. N. and McNabb, W. C., 1999. The implications of condensed tannins on the nutritive value of temperate forages fed to ruminants. *Br. J. Nutr.* **81**: 263-272.

Barry, T. N., McNeill, D. M., and McNabb, W. C., 2001. Plant secondary compounds; their impact on forage nutritive value and upon animal production. *Grassl. Conf. Sao Paulo, Brazil*. 445-452.

Barry, T.N. and Forss, D., 1983. The condensed tannin content of vegetative *Lotus pedunculatus*, its regulation by fertiliser application, and effect upon protein solubility. *Journal of Science and Food in Agriculture*. **34**: 1047-1056.

Barry, T.N., Manley, T.R. and Duncan, S.J., 1986. The role of condensed tannins in the nutritional value of *Lotus pedunculatus* for sheep. 4-Sites of carbohydrate and protein digestion as influenced by dietary reactive tannin concentration. *British Journal of Nutrition*. **55**: 123-137.

Beever, D .E., Terry, R. A., Cammell, S.B, Wallace, A.S., 1978. The digestion of spring and autumn harvested perennial rye grass by sheep. *Journal of Agricultural Science Cambridge*. **90**: 463-470.

Behnke, R.H. and Scoones, I.,1993. Rethinking range ecology: Implications for rangeland management in Africa. *Range ecology at disequilibrium*, Overseas Development Institute, London.pp.1-30.

Ben Salem, H., Nefzaoui, A., Makkar, H.P.S., Hochlef, H., Ben Salem, I. and Ben Salem, L., 2005. Effect of early experience and adaptation period on voluntary intake, digestion and growth in Barbarine lambs given tannin-containing (*Acacia cyanophylla* Lindl. foliage) or tannin-free (oaten hay) diets. *Anim. Feed Sci. Technol.* **122**: 59-77.

- Berhanu, G, Adane, H. and Kahsay, B., 2009. Feed marketing in Ethiopia: Results of rapid market appraisal. Working Paper No. 15. ILRI (International Livestock Research Institute), Addis Ababa, Ethiopia.
- Bogdan, A. V., 1977. Tropical pasture and fodder plants. Longman. pp. 475.
- Bullis, D.D., Haj-Manouchehri, M.A. and Know, K.L., 1967. In situ nylon bags dry matter digestibility as a predictor of ration feeding value. *J Anim Sci.* **26**:130.
- Butler, L.G., 1988. The role of polyphenols in the utilization of ICRISAT-mandated grain crops and applications of biotechnology for improved utilization. In Biotechnology in tropical crop improvement. *Proceedings of the International Biotechnology Workshop.* 147-152.
- Cabrita, A. R. J.; Dewhurst, R. J.; Abreu, J. M. F. and Fonseca, A. J. M., 2006. Evaluation of the effects of synchronizing the availability of N and energy on rumen function and production responses of dairy cows. *Animal Research.* **55**:1-24.
- Carlos, A., Sandoval, C., Henry, L., Lizarraga, S., Francisco, J. and Solorio, S., 2005. Assessment of tree fodder preference by cattle using chemical composition, *in vitro* gas production and *in situ* degradability. *Anim. Feed Sci. Technol.* pp. 123-124, 277-289.
- Cassida, K.A., Griffin, T.S., Rodriguez, J., Patching, S.C., Hesterman, O.B., and S. R. Rust., 2000. Protein degradability and forage quality in maturing alfalfa, red clover, and birdsfoot trefoil. *Crop Sci.* **40**:209-215.
- Caton, J.S., Freeman, A.S. and Galyean, M.L., 1988. Influence of protein supplementation on forage intake, in situ forage disappearance, ruminal fermentation and digesta passage rates in steers grazing dormant blue grama rangeland. *J Anim Sci.* **66**: 2262-2271.
- Chang, S. I. and Fuller, H. L., 1964. Effect of Tannin content of grain sorghum on their feeding value for growing chicks. *Poultry Science.* **43**: 30-37.
- Chaudhry, A.S. and Webster, A.J.F., 2001. Nutrient composition and the use of solubility to estimate rumen degradability of food proteins in cattle. *J Sci Food Agric.* **81**: 1077-1086.
- Chaudhry, A.S., 2007. Enzymic and in sacco methods to estimate rumen degradation of food protein in cattle. *J Sci Food Agric.* **26**: 2617-2624.
- Cheema, U. B.; Sultan, J. I.; Javaid, A.; Akhtar, P. and Shahid, M., 2011. Chemical composition, mineral profile and *in situ* digestion kinetics of fodder leaves of four native trees. *Pakistan J. Bot.* **43 (1)**: 397-404.
- Cheng, K.J., Stewart, C.S., Dinsdale, D, and Costerton, J.W., 1984. Electron-microscopy of bacteria involved in the digestion of plant-cell walls. *Anim Feed Sci Technol.* **10**: 93-120.

Cockburn, J.E., Dhanoa, M.S., France, J. and Lopez, S., 1993. Overestimation of solubility when using Dacron bag methodology. In Proceedings of the British Society of Animal Science, Scarborough, p. 188. Penicuik, Scotland: British Society of Animal Science.

D'Mello, J.P.F., 1982. Toxic factors in some tropical legumes. *World Review Animal Production*. **18**: 41-46.

De Leeuw, P.N. and Rey, B., 1995. An analysis of the current trends in the distribution of Ruminant livestock in the Sub-Saharan Africa. *World Animal Review* **83**: 43-59.

De Smet, A.M., De Boever, J.L., De Brabander, D.L., Vanacher, J.M. and Boucque, V.C., 1995. Investigation of dry matter degradation and acidotic effect of some feedstuffs by means of in sacco and in vitro incubation. *Anim Feed Sci Technol* **51**: 297-315.

Decruyenaere, V., Lecomte, P.H., Demarquilly, C., Aufrere, J., Dardenne, P., Stilmant, D. and Buldgen, A. 2009. Evaluation of green forage intake and digestibility in ruminants using near infrared reflectance spectroscopy (NIRS): Developing a global calibration. *Anim. Feed Sci Tech.* **148**: 138- 156.

Devendra, C., 1989. Ruminant production systems in the developing countries: resource utilisation, In Feeding Strategies for Improved Productivity of Ruminant Livestock in Developing Countries, IAEA, Vienna, Austria. pp 5-30.

Devendra, C., 1995. Composition and nutritive value of browse legumes. In: (Ed. J. P. F. D'Mello and C. Devendra). Tropical Legumes in Animal Nutrition. CAB International, Wallingford, UK. pp 49-65.

D'Mello and C. Devendra). Tropical Legumes in Animal Nutrition. CAB International, Wallingford, UK. pp 49-65.

Djouvinov, D.S., Nakashima, Y., Todorov, N. and Pavlov, D., 1998. In situ degradation of feed purines. *Anim Feed Sci Technol.* **71**: 67-77.

El-hassan, S.M., Lahlou, K.A, Newbold, C.J. and Wallace, R.J., 2000. Chemical composition and degradation characteristics of some African Multipurpose trees. *Ann. Feed Sci. Technol.* **86**: 27-37.

Fadiyimu, A.A., Fajemisin, A.N., Arigbede, M.O. and Alokun, J.A., 2012. Rumen Dry Matter Degradability and preference by West African Dwarf Goats for Selected Multipurpose Trees in Nigeria. *Livestock Research for Rural Development.* **24(1)**.

Fahey, Jr., G.C. and Jung, H.G., 1989. Phenolic compounds in forage and fibrous feedstuffs. Toxicants of plant origin. In: Cheeke, P.R. (Ed.), Phenolics, vol. IV. CRC Press, Inc., Boca Raton, FL. pp. 123-190.

FAO/IAEA., 2000. Quantification of Tannins in Tree Foliage. IAEA Working document, IAEA, Vienna.

- Ferreira, A.V., 2004. Nutritive value of red vine husks and pips for sheep. *South African Journal of Animal Science*. **34 (2)**:23-25.
- Figtoid, W., Hale, W. H. and Theurer, B., 1972. An evaluation of the nylon bag technique for estimating rumen utilization of grains. *J. Anim. Sci.* **35**: 113.
- Fraser, M.D., Theobald, V.J., Davies, D.R. and Moorby, J.M., 2009. Impact of diet selected by cattle and sheep grazing heathland communities on nutrient supply and faecal micro-flora activity. *Agric Ecosystems and Environment*. **129**: 367-377.
- Frutos, P., Hervas, G., Giraldez, F.J., Fernandez, M. and Manteco, A.R., 2000. Digestive utilization of quebracho-treated soya bean meal in sheep. *J Agr Sci.* **134**: 101-108.
- Gasmi-Boubaker, A., Kayouli, C. and Buldgen, A., 2005. In-vitro gas production and its relationship to in situ disappearance and chemical composition of some Mediterranean browse species. *Animal Feed Science and Technology*. **123(1)**: 303-311.
- Gerhard, F. and Peter, L., 1997. Developmental changes of carbohydrates in Cape gooseberry (*physalis peruviana* L.) fruits in relation to the calyx and the leaves. *Agronomia colombiana*. **14 (2)**: 95-107.
- Getachew, G., Makkar, H. P. S. and Becker, K., 2000. Effect of polyethylene glycol on in vitro degradability of nitrogen and microbial protein synthesis from tannin-rich browse and herbaceous legumes. *British Journal of Nutrition*. **84**: 73-83.
- Giang, H. H., Ly, L. V. and Ogle, B., 2004. Evaluation of ensiling methods to preserve sweet potato roots and vines as pig feed. *Livestock Research for Rural Development*. **16 (7)**. http://www.Cipav.org.co/lrrd/lrrd/16/7/gian16_045.htm
- Gillespie, J. R., 1998. *Animal Science*. Delmar publishers, International Thompson Publishing Company, pp 1204.
- Gomes, H. S., 1990. Phenolic profile of Shrub Live Oak and its relation to goat diets in Central Arizona. Ph. D. Dissertation. University of Arizona, Tucson.
- Gonzalez, J., Santiago, A., Carlos, A. R. Maria, R. A., 2002. In situ evaluation of the protein value of soybean meal and processed full fat soybeans for ruminants *Anim.Ref.* **51(6)**: 455-464
- GraphPad, Prism., 2009. Version 5 for Windows. 2009. GraphPad Software, San Diego, CA, USA.
- Guerin, H., 1987. Alimentation des ruminants domestiques sur pâturages naturels sahéliens et Gupta, P., Kalpana, S. and Avinash.U., 2011. Isolation of Cellulose-Degrading Bacteria and Determination of Their Cellulolytic Potential. *International Journal of Microbiology*.

Hagerman, A.E. and Butler, L.G., 1981. The specificity of proanthocyanidin-protein interactions. *Journal of Biological Chemistry*. **256**: 4494-7.

Hagerman, A.E., 2011. The tannin handbook. USDA (United States Department of Agriculture). <http://www.users.muohio.edu/hagermae>.

Haslam, E., 1989. Plant Polyphenols. Vegetable Tannins Revisited. Cambridge University Press, Cambridge, UK.

Hervas, G., Frutos, P., Serrano, E., Manteco, A. R.N. and Gira, F. J., 2000. Effect of tannic acid on rumen degradation and intestinal digestion of treated soya bean meals in sheep. *Journal of Agricultural Science, Cambridge*. **135**: 305-310.

Hoang, H. G.; Le, V.L. and Ogle, B., 2004. Evaluation of ensiling methods to preserve sweet potato roots and vines as pig feed. *Livestock Research for Rural Development*. **16 (7)**: 45-46.

Holechek, J.L., Vavra, M. and Pieper, R.D., 1982. Methods for determining the nutritive quality of range ruminant diets: A review. *J Anim Sci*. **54**: 363-376.

Huntington, J.A. and Givens, D.I., 1995. The in situ technique for studying rumen degradation of feeds. A review of the procedure. *Nutr Abstr Rev*. **65(2)**: 63-93.

Hvelplund, T. and Weisbjerg, M.R., 1998. In vitro techniques to replace in vivo methods for estimating amino acid supply. Penicuik, UK. *British Society of Animal Science*. **22**: 131-144.

Ibrahim, H., 1999. Feed Resources for Ruminant Livestock. ILRI Slide Series 1. ILRI.

Ikhimioya, I., 2008. Acceptability of selected common shrubs/trees leaves in Nigeria by West African dwarf goats. *Livestock Research for Rural Development*. **20 (6)**.

Ikhimioya, I., Isah, O.A., Ikhatua, U.J. and Bamikole, M.A., 2005. Rumen Degradability of Dry Matter and Crude Protein in Tree Leaves and Crop Residues of Humid Nigeria. *Pakistan Journal of Nutrition*. **4 (5)**: 313-320.

Jolly, S. and Wallace, A., 2007. Best practice for production feeding of lambs: a review of the literature. Feedlot mid North SA (Productive Nutrition Pty Ltd).

Jones, W.T., Anerson, L.B. and Ross, M.D., 1973. Bloat in cattle, Detection of protein precipitants (flavolans) in legumes. *New Zealand Journal of Agricultural Research*. **16**: 441-446.

Kabuga, J.D. and Darko, C.A., 1993. *In sacco* degradation of dry matter and nitrogen in oven dried and fresh tropical grasses and some relationships to in vitro dry matter digestibility. *Animal Feed Science and Technology*. **40**:191-205.

Kadzere, C.T., 1995. Feed resources for sustainable ruminant livestock production in southern Africa. *Afr. Stud. Monogr*. **16(4)**: 165

- Kaitho, R. J.; Tegegne, A.; Umunna, N. N.; Nsahlai, I. V.; Tamminga, S.; Bruchem, J.V. and Arts, J. M., 1998. Effect of Leucaena and Sesbania supplementation on body growth and scrotal circumference of Ethiopian highland sheep and goats fed teff straw basal diet. *Livest. Prod. Sci.* **54** (2): 173-181.
- Kanani, J.; Lukefahr, S. D. and Stanko, R. L., 2006. Evaluation of tropical forage legumes (*Medicago sativa*, *Dolichos lablab*, *Leucaena leucocephala* and *Desmanthus bicornutus*) for growing goats. *Small Ruminant Research.* **65**: 1-7.
- Kechero, Y. and Duguma, B., 2011. Evaluation of Nutritive Value of *Albizia gummifera* Foliages as Fodder Source for Livestock in Agrisilvipastoral System. *International Journal of Agricultural Research.* **6**: 389-399.
- Kendall, W. A., 1966. Factors affecting foams with forage legumes. *Crop Sci.* **6**:487-489.
- Khatab, R.Y. and Arntfield, S.D., 2009. Nutritional quality of legume seeds as affected by some physical treatments 2. Antinutritional factors. *Food Sci. Tech.* **42**: 1113- 1118.
- Kibon, A., Orskov, E.R., 1993. The use of degradation characteristics of browse plants to predict intake and digestibility by goats. *Anim. Prod.* **57**: 247–251.
- Kindu, M., Glatzel, G. and Sieghardt, M., 2006. Evaluation of common indigenous tree and shrub species for soil fertility improvement and fodder production in the highland areas of western Shewa, Ethiopia. PP. 99-106.
- Krishnamoorthy, U.K., Rymer, C. and Robinson, P.H., 2005. The in vitro gas production technique: limitation and opportunities. *Anim Feed Sci Technol.* 123-124.
- Kristensen, E.S, Moller, P.D and Hvelplund, T., 1982. Estimation of the effective protein degradability in the rumen of cows using nylon bag technique combined with outflow rate. *Acta Agric Scand.* **32**: 123-127.
- Kumar, R. and Sing, M., 1984. Tannins: their adverse role in ruminant nutrition. *J. Agric. Food Chem.* **32**: 447-453.
- Kumar, R. and Vaithyanathan, S., 1990. Occurrence, nutritional significance and effect on animal productivity of tannins in tree leaves. *Animal Feed Science and Technology.* **30**: 21-28.
- Lamers, J. P. A. and Khamzina, A., 2010. Seasonal quality profile and production of foliage from trees grown on degraded cropland in arid Uzbekistan, Central Asia. *Journal of Animal Physiology and Animal Nutrition.* **94**: 77-85.

- Landau, S., Silanikove, N., Nitsan, Z., Barkai, D., Baram, H., Provenza, F.D. and Perevolotsky, A., 2000. Short – term changes in eating patterns explain the effects of condensed tannins on feed intake in heifers. *Applied animal Behavior Science*. **69**: 199-213.
- Lanyasunya, T. P.; Wang, H. R.; Mukisira E. A.; Abdulrazak, S. A. And Ayako, W. O., 2006. Effect of seasonality on feed availability, quality and herd performance on smallholder farms in ol-joro-orok location/nyandarua district. *Tropical and Subtropical Agroecosystems*. **6 (2)**: 87-93.
- Larbi, A., Smith, J.W., Kurdi, I.O., Adekunle, I.O, Raji, A.M. and Ladipo, D.O., 1998. Chemical composition, rumen degradation, and gas production characteristics of some multipurpose fodder trees and shrubs during wet and dry seasons in the humid tropics. *Anim. Feed Sci. Technol.* **72**: 81-96.
- Larbi, A., Smith, J.W., Raji, A.M., Kurdi, I.O., Adekunle, I.O. and Ladipo, D.O., 1997. Seasonal dynamics in dry matter degradation of browse in cattle, sheep and goats. *Small Rumin Res.* **25**:129-140.
- Le Houerou, H.N., 1980. Browse in Africa. In: Proceedings of the International Symposium International Livestock Centre for Africa (ILCA). Addis Ababa. P 491.
- Lebopa, C.K.; Boomker, E.A.; Chimonyo, M. and Mulugeta, S.D., 2011. Factors affecting the feeding behaviour of free ranging Tswana and Boer goats in the False Thornveld of the Eastern Cape, South Africa. *Life Science Journal*. **8(2)**.
- Lees, G. L., Suttill, N. H. and Gruber, M. Y., 1993. Condensed tannins in sainfoin I. A histological and cytological survey of plant tissues. *Can. J. Bot.* **71**:1147–1152.
- Leinmuller, E., Steingass, H. and Menke, K.H., 1991. Tannins in ruminant feedstuffs. Biannual Collection of Recent German Contributions Concerning Development through Animal Research **33**: 9-62.
- Lopez, S., Hovell, F.D., Manyuchi, B. and Smart, I., 1995. Comparison of sample preparation methods for the determination of the rumen degradation characteristics of fresh and ensiled forage by nylon bag technique. *Anim Sci.* **60**: 439-450.
- Lowry, J.B., McSweeney, C.S. and Palmer, B., 1996. Changing perceptions of the effect of plant phenolics on nutrient supply in the ruminant. *Aust. J. Agric. Res.* **47**: 829-842.
- Ly, J., Pok, S. and Preston, T.R., 2001. Nutritional evaluation of tropical leaves for pigs: Pepsin/pancreatin digestibility of thirteen plant species. *Livest. Res. Rural Dev.* **13(5)**.
- Madsen, J. and Hvelplund, T., 1994. Prediction of in situ protein degradability in the rumen. Results of a European ring test. *Livest Prod Sci.* **39**: 201-212.

Mahadevan, S., Erfle, J.D. and Sauer, F.D., 1980. Degradation of soluble and insoluble proteins by *Bacteroides amylophilus* protease and by rumen microorganisms. *J. Anim. Sci.* **50**: 723-728.

Makkar, H. P. S., 1993. Antinutritional factors in foods for livestock. In: *Animal Production in Developing Countries*, Occasional Publication No 16, British Society of Animal Production., ed Gill M, Owen E, Pollott G E and Lawrence T L J. pp. 69-85.

Makkar, H. P. S., 2003. Effects and fate of tannins in ruminant animals, adaptation to tannins, and strategies to overcome detrimental effects of feeding tannin-rich feeds. *Small Rum. Res.* **49**: 241-256.

Makkar, H.P.S. and B. Singh., 1991b. Distribution of condensed tannins (proanthocyanidins). In various fractions of young and mature leaves of some oak species. *Anim. Feed Sci. Technol.* **32**: 253-260.

Makkar, H.P.S. Becker, K., 1997. Potential of *Jatropha* seed cake as protein supplement in livestock feed and constraints to its utilization. In: *Proc Jatropha 97: International symposium on Biofuel and Industrial Products from Jatropha curcas and other T tropical Oil Seed Plants*. Managua/ Nicaragua, Mexico, 23E27 Feb 1997.

Makkar, H.P.S., Borowy, N.K., Becker, K. and Degen, A., 1995. Some problems in fiber determination of a tannin – rich forage (*Acacia saligna* leaves) and their implications in *in vivo* studies. *Animal Feed Science and Technology.* **55**: 67 -76.

Makkar, H.P.S., Singh, B. and Kamra, D.N., 1994. Biodegradation of tannin in oak (*Quercus incana*) leaves by *Sporotricum pulverulentum*. *Lett. Appl. Microbiol.* **18**:42-44.

Makkar, H.P.S., Singh, B. and Negi, S.S., 1989. Relationship of rumen degradability with microbial colonization, cell wall constituents and tannin levels in some tree leaves. *Anim. Prod.* **49**: 299-303.

Mandal, L. 1997. Nutritive values of tree leaves of some tropical species for goats. *Small Rumin.*

Mangan, J., 1988. Nutritional effects of tannins in animal feeds. *Nutrition Research Reviews.* **1**: 209-231.

Marinucci, MT., Dehority, B.A. and Loerch, S.C., 1992. In vitro and in vivo studies of factors affecting digestion of feeds in synthetic fiber bags. *J Anim Sci.* **70**: 296–307.

Mc Nabb, W.C., Waghorn, G.C., Peters, J.S. and Barry, T.N., 1996. The effect of condensed tannins in *Lotus pedunculatus* on the stabilization and degradation of ribulose-1,5-bisphosphate carboxylase (EC 4.1.1.39: rubisco) protein in the rumen and the sites of rubisco digestion. *Br. J. Nutr.* **76**: 535-549.

McDonald, P., Edwards, R.A, Greenhalgh, J.F.D. and Morgan, C.A., 2002. *Animal Nutrition*. Pearson Education Limited, sixth edition, London.

- McLeod, M.N., 1974. Plant tannins - Their role in forage quality. *Nutr Abst Rev.* **44**: 803-812.
- McMahon, L.R., McAllister, T.A., Berg, B.P., MA- Jak, W., Acharya, S.N., Popp, J.D., Coulman, B.E., Wang, Y. and Cheng, K.J., 2000. A review of the effects of forage condensed tannins on ruminal fermentation and bloat in grazing cattle. *Can J Plant Sci.* **80**, 469-485.
- McNaughton, S.J., 1987. Adaptation of herbivores to season changes in nutrient supply. In: Nutrition of Herbivores (Hecker, J.B and Ternouth, J.H., eds.). *Academic Press, Sydney, Australia.* pp. 391-408.
- McSweeney, C.S., Kennedy, P.M. and John, A., 1988. Effect of ingestion of hydrolysable tannins in *Terminalia oblongata* on digestion in sheep fed *Stylosanthes hamata*. *Australian Journal of Agricultural Research.* **39**: 235-244.
- Mehrez, A.Z. and Orskov, E.R., 1977. A study of the artificial fiber bag technique for determining the digestibility of feeds in the rumen. *J Agric Sci Camb.* **88**: 645-650.
- Meissner, H.H., Viljoen, M.O. and Van Niekerk, W.A., 1991. Intake and digestibility by sheep of Antherphora, Panicum, Rhodes and Smooth finger grass In: Proceedings of the IVth International Rangeland Congress, September 1991, Montpellier, France. pp 648-649.
- Mekoya, A., Oosting, S.J., Fernandez-Rivera, S. and Van der Zijpp, A.J., 2008. Multipurpose fodder trees in the Ethiopian highlands: Farmers' preference and relationship of indigenous knowledge of feed value with laboratory indicators. *Agricultural Systems.* **96**: 184-194.
- Menke, K.H., Raab, L., Salewski, A., Steingass, H., Fritz, D. and Schneider, W., 1979. The estimation of the digestibility and metabolisable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor in vitro. *J Agric Sci.* **93**: 217-222.
- Mertens, D.R., 1993. Kinetics of cell wall digestion and passage ruminants. In: Jung HG, Buxton DR, Hatfield RD, Ralph, editors. Forage cell wall structure and digestibility. USDA, Agricultural Research Service and US Dairy Forage Research Center, Madison, WI. pp 538-570.
- Meuret, M., Boza J., Narjisse, N. and Nastis, A., 1990. Evaluation and utilization of rangeland feeds by goats. In: Goat Nutrition, PUDOC. (Ed.): P. Morand-Fehr. Wageningen. *The Netherlands.* pp 161-170.
- Meyer, J.H.F. and Mackie, R.I., 1986. Microbiological evaluation of the intraruminal in sacco digestion technique. *Appl Environ Microbiol.* **51**: 622-634.
- Mezzomo, R., Michalet-Doreau, B. and Cerneau, P., 1991. Influence of foodstuff particle size on in situ degradation of nitrogen in the rumen. *Anim. Feed Sci. Technol.* **35**: 69-81.

Michalet-Doreau, B., and Champion, M., 1995. Influence of maize genotype on rate of ruminal starch degradation. *Ann. Zootech.* 44(Suppl. 1):191 (Abstr.).

Migongo-Bake, W., 1992. Rumen dry-matter digestive efficiency of camels, cattle, sheep and goats in a semiarid environment in Eastern Africa. International Livestock Center for Africa (ILCA), Addis Ababa, Ethiopia. pp. 27-35.

Min B.R., McNabb W.C., Barry T.N., Kemp P.D., Waghorn G.C., McDonald M.F., 1999. The effect of condensed tannins in *Lotus corniculatus* upon reproductive efficiency and wool production in sheep during late summer and autumn. *J Agr Sci.* **132**: 323-334.

Min, B.R., Hart S.P., 2003. Tannins for suppression of internal parasites. *J Anim Sc* 81, E. Suppl. 2, E102-E109.

MoA(Ministry of Agriculture)., 1997. Ministry of Agriculture, National Livestock Development Programme (NLDP). Main working Papers 1-3, Addis Ababa, Ethiopia.

Morris, P., Carron, T. R., Robbins, M. P. and Webb, K. J., 1993. Distribution of condensed tannins in flowering plants of *Lotus corniculatus* var. japonicas and tannin accumulation by transformed root cultures. *Lotus Newsletter.* 24: 60-63.

Mueller-Harvey, I., 2006: Unravelling the conundrum of tannins in animal nutrition and health. *Journal of the Science of Food and Agriculture.* **86**: 2010-2037.

Mupangwa, J. F., Ngongoni, N. T., Topps, J. H., Acamovic, T., Hamudikuwanda, H., 2003. Rumen degradability and post-ruminal digestion of dry matter, nitrogen and amino acids in three tropical forage legumes estimated by the mobile nylon bag technique. *Livestock Prod. Sci.* **79(1)**: 37-46.

Nashlai, I.V., Siaw ,D.E.K. and Osuji, O.P., 1994. The relationship between gas production and chemical composition of 23 browses of the genus *Sasbania*. *J. Sci. Food. Agric.* **65**: 13-20.

Nasrullah, M.N., Akhasi, R. and Kawamure, O., 2003. Nutritive value of forage plants grown in South Sulawesi, Indonesia. *Asian-Aust. J. Anim. Sci.* **16**: 693-701.

National Research Council., 1981. Nutrient requirements for goats Angora, dairy and meat goat in temperate and tropical countries. National Research Council. National Academy of Science Press, Washington D.C. U.S.A.

National Research Council., 1996. Nutrient Requirements for Beef Cattle, 7th ed. Washington, DC: National Academy Press.

National Research Council., 2001. Nutrient Requirements for Dairy Cattle, 7th ed. National Academy Press, Washington D.C. U.S.A.

Ngodigha, E.M and Oji, U.I., 2009. Evaluation of fodder potential of some tropical browse plants using fistulated N'dama cattle. *African Journal of Agricultural Research*. **4(3)**: 241-246.

Ngodigha, E.M. and Anyanwu, N.J., 2009. Fodder Potential Ranking of Selected Multi-Purpose Trees and Shrubs Through Degradation Studies with Rumen Fistulated N'dama Steers. *Journal of Animal and Veterinary Advances*. **8(6)**: 1233-1236.

Nocek, J.E., 1988. In situ and other methods to estimate ruminal protein and energy digestibility. A review. *J Dairy Sci*. **71**: 2051–2069.

Nordblom, J.L. and Shomo, F., 1995. Food and Feed Prospects to 2020 in the West Asia North Africa Region. ICARDA Social Science Paper No. 2, International Centre for Agricultural Research in the Dry Areas, Aleppo, Syria.

Norton, B.W 2003. The Nutritive value of tree legumes. In: Forage tree legumes in tropical agriculture (Ed. R. C. Gutteridge and H.M. Shelton). pp 1-10.
Web site: <http://www.fao.org/ag/AGP/AGPC/doc/Publicat/Gutt-shel/x5556e0j.htm>.

Norton, B.W., 1994. Tree legumes as dietary supplements for ruminants. In: Gutteridge, R.C. and H.M. Shelton (editors). Forage tree legumes in tropical agriculture, CAB International. pp 192-201.

Norton, B.W., 2000. The significance of tannins in tropical agriculture. In: Brooker, J.O. (Ed). Tannins in Livestock and Human Nutrition. Proceedings of an International Work shop, Adelaide, Australia, May 31-June 2, 1999. Canberra. ACIAR (Australian Centre for International Agricultural Research) proceedings No. 92. pp. 14-23.

Noziere, P. and Michalet-Doreau, B., 1996. Validation of in sacco method: influence of sampling site, nylon bag or rumen contents, on fibrolytic activity of solid-associated microorganisms. *Anim Feed Sci Technol*. **57**: 203-210.

Noziere, P. and Michalet-Doreau, B., 2000. In sacco methods. In Farm Animal Metabolism and Nutrition. pp. 233–254.

Odedire, J. A.; Babayemi, O. J., 2008. Comparative studies on the yield and chemical composition of Panicum maximum and Andropogon gayanus as influenced by Tephrosia candida and Leucaena leucocephala. *Livestock Research for Rural Development*. **20(2)**.
<http://www.lrrd.org/lrrd20/2/oded20027.htm>

Ondiek, J. O., Abdulrazak, S. A., Njoka, E. N., 2010. Chemical and mineral composition, *in-vitro* gas production, *in-sacco* degradation of selected indigenous Kenyan browses. *Livestock Research for Rural Development*. **22(2)**: 25.

Onwuka, C. F. I., 1992: Tannin and saponin contents of some tropical browse species fed to goats. *Tropical Agriculture Trinidad*. **69**: 177-180.

- Orskov, E. R. and McDonald, I., 1979. The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *Journal of Agricultural Science (Cambridge)*. **92**: 499-503.
- Orskov, E.R., Hovell, F.D., and Mould. F., 1980. The use of the nylon bag technique for the evaluation of feedstuffs. *Tropical Animal Production*. **5**: 195 - 213.
- Orskov, E.R., Oiwang, I. and Reid, G.W., 1988. A study on consistency of differences between cows in rumen outflow rate of fibrous particles and other substrates and consequences for digestibility and intake of roughages. *Animal Production*. **47**: 45-51.
- Osuji, P. O., Nsahlai, I.V. and Khalili, H., 1993. Feed evaluation. ILCA Manual 5. ILCA (International Livestock Centre for Africa), Addis Ababa, Ethiopia. pp40.
- Owens, F. N.; Secrist, D. S.; Hill, W. J. and Gill, D. R., 1998. Acidosis in cattle: a review. *Journal of Animal Science*. **76**: 275-286.
- Ozkan, C. O. and M. Sahin., 2006. Comparison of *In situ* Dry Matter Degradation with *In vitro* Gas Production of Oak Leaves Supplemented with or without Polyethylene Glycol (PEG). *Asian-Aust. J. Anim. Sci.* **19(8)**: 1120 – 1126.
- Ozturk, D., Ozkan C.O., Atalay A.I., Kamalak A., 2006. The Effect of Species and Site on the Condensed Tannin Content of Shrub and Tree Leaves. *Research Journal of Animal and Veterinary Sciences*. **1(1)**: 41-44.
- P.V.R. Paulino, E., Detmann, S.C., Valadares, F., Paulino, M.F., Monnerat, J.P.I.S., Duarte, M.S., Silva, L.H.P. and Moura, L.S., 2011. *Livestock Science*. **141(1)**: 1-11.
- Papachristou, T.G., Papanastasis, V. P., 1994. Forage value of Mediterranean deciduous woodyfodder species and its implication to management of silvo-pastoral systems for goats. *Agroforestry Systems*. **27**: 269-282.
- Patra, A. K., 2010: Effects of supplementing low-quality roughages with tree foliages on digestibility, nitrogen utilization and rumen characteristics in sheep: a meta-analysis. *Journal of Animal Physiology and Animal Nutrition*. **94**: 338-353.
- Perevolotsky, A., A. Brosh. O., Ehrlich, M., Gutnian, Z. and Holtezer, Z., 1993. Nutritional values of common oak (*Quercus calliprinos*) browse as fodder for goats: Experimental results inecological perspective. *Small Rumin. Res.* **11**: 95-106.
- Playne, M. J., Khumnualthong, W. and Echevarria, M. G., 1978. Factors affecting the digestion of oesophageal fistula samples and hay samples in nylon bags in the rumen of cattle. *J. Agr. Sci.* **90** :193.
- Polshettiwar, S.A., R.O. Ganjiwale, S.J., Wadher, P.G., 2007. Spectrophotometric estimation of

total tannins in some ayurvedic eye drops. *J. Pharm.Sci.* **69**: 574-576.

Preston, T.R., 1995. Tropical animal feeding. A manual for research workers. FAO Animal Production and Health Paper 126. FAO. Rome.

Web site: <http://www.fao.org/DOCREP/003/V9327E/V9327E00.HTM>

Proietti, P., Palliotti, A., Famiani, F., Antognozzi, E., Ferranti, F., Andreutti, R. and Frenguelli, G., 2000. Influence of leaf position, fruit and light availability on photosynthesis of two chestnut genotypes. *Sci. Hortic.* **85**: 63-73.

Provenza, F. D., Lynch, J. J. and Cheney, C. D., 1995. Effects of a flavor and food restriction on the intake of novel foods by sheep. *Appl. Anim. Behav. Sci.* **43**:83.

Provenza, F.D., 1995. Postingestive feedback as an elementary determination of food selection and intake in ruminants. *Journal of Range Management.* **48**: 2 -17.

Provenza, F.D., Burritt, E.A., Clausen, T.P., Bryant, P.B. & Distel, R.A., 1990. Food Aversion Learning in Goats: A Mechanism to Avoid Condensed Tannins in Blackbrush. *American naturalist.* **136**: 810-828.

Quin, J.I, Van der Wath, J.G. and Myburgh, S., 1938. Studies on the alimentary tract of merino sheep in South Africa. Part IV. Description of experimental technique. *J Vet Sci Anim Ind.* **11**: 341-361.

Ramana, D.B.V., Sultan Singh, K.R. and Solanki, A.S., 2000. Nutritive evaluation of some nitrogen and non-nitrogen fixing multipurpose tree species. *Animal Feed Science and Technology.* **88**:103-111.in

Ramirez, R.G., Neira-Morales, R.R., Ledezma-Torres, R.A. and Garibaldi-Gonzalez, C.A., 2000. Ruminant digestion characteristics and effective degradability of cell wall of browse species from north eastern Mexico. *Small Rumin Res.* **36**: 49-55.

Reed, J. and Soller, H., 1987. Phenolics and nitrogen utilization in sheep fed browse. In: M. Rose (eds), *Herbivore Nutrition Research, 2nd International Symposium on the Nutrition of Herbivores*, University of Queensland, Brisbane. pp. 47-8

Reed, J. D., 1995. Nutritional toxicology of tannins and related polyphenols in forage legumes. *J. Anim. Sci.* **73**:1516-1528.

Robbins, C.T., Hanely, T.A., Hagerman, A.E., Hjeljord, O., Baker, D.L., Schwartz, C.C. & Mautz, W.W., 1987. The role of tannins in defending plants against ruminant reduction in protein availability. *Ecology.* **68**: 98-07.

Rubanza, C.D.K., Shem, M.N, Otsyina, R., Bakengesa, S.S., Ichinohe, T. and Fujihara, T., 2005. Polyphenolics and tannins effect on *in vitro* digestibility of selected Acacia species leaves. *Animal Feed Science and Technology.* **119**: 129-142.

sahelo - soudaniens: etude methodologique dans la region du Ferlo au Senegal. These Doct.,

Salem, A.Z.M., Salem, M.Z.M., El-Adawy, M.M. and Robinson, P.H., 2006. Nutritive evaluations of some browse tree foliages during the dry season: Secondary compounds, feed intake and *in vivo* digestibility in sheep and goats. *Anim. Feed Sci. Technol.* **127**: 251-267.

Salunkhe, D.K., Chavan, J.K. and Kadam, S.S., 1990. Dietary Tannins: Consequences and Remedies. Boca Raton, FL, CRC Press.

Schlink, A. C.; Burt, R. L., 1993. Assessment of the chemical composition of selected tropical legume seeds as animal feed. *Trop. Agric. Trinid.* **70**: 169-173.

Schmidk, A., Takahashi, R., De Medeiros, A.N. and De Resende, K.T., 2000. Bromatological composition and degradation rate of mulberry in goats. In: *FAO Electronic Conference on Mulberry for Animal Production (Morus1-L)*;
<http://www.fao.org/ag/AGA/AGAP/FRG/Mulberry/Posters/PDF/Schmidk1>.

Silanikove, N., Gilboa, N., Nir, I., Perevolotsky, A. and Nitsan, Z., 1996. Effect of a daily supplementation of polyethylene glycol on intake and digestion of tannin-containing leaves (*Quercus calliprinos*, *Pistacia lentiscus*, and *Ceratonia siliqua*) by goats. *Journal of Agricultural Food Chemistry.* **44**: 199-205.

Silanikove, N., Nitsan, Z. and Perelovsky, A., 1994. Effect of a daily supplementation of polyethylene glycol on intake and digestion of tannin-containing leaves (*Ceratonia siliqua*) by sheep. *J. Agric. Food Chem.* **42**:2844-2847.

Silanikove, N., Nitsan, Z. and Perevolotsky, A., 1994. Effect of a daily supplement of polyethylene glycol on intake and digestion on tannin-containing leaves (*Ceratonia siliqua*) by sheep. *J Agric Food Chem.* **42**: 2844 - 2847.

Silanikove, N.; Gilboa, A.; Nitsan, Z. and Perevolotsky, A., 1997. Effect of foliage-tannins on feeding in goats. *Options Me'diterrane'ennes.* **34 (1)**: 43-45.

Silanikove, N.; Perevolotsky, A. And Provenza, F. D., 2001. Use of tannin-binding chemicals to assay for tannins and their negative postingestive effects in ruminants. *Anim. Feed Sci. Technol.* **91 (1/2)**: 69-81.

Singh, B, Makkar, H.P.S. and Negi, S.S., 1989. Rate and extent of digestion and potentially digestible dry matter and cell wall of various tree leaves. *J Dairy Sci.* **72 (12)**: 3233-3239.

Singleton, D. L.M., 1981. Naturally occurring food toxicants: phenolic substance of plant origin common in woods. *Advanced in Food Research.* **27**: 149.

Skerman, P. J. and Riveros, F., 1990. Tropical Grasses. Food and Agric. Org. United Nations; Plant Prod. Protect. No. 23, Rome, Italy.

Smith, O.B., 1992. Fodder trees and fodder shrubs in range and farming systems of tropical humid Africa. In: Speedy, A. and Pugliese, P.L. (Eds), Legume trees and other fodder trees as protein sources for livestock. FAO (Food and Agriculture Organization) Animal Production and Health Paper No.102. FAO, Rome, Italy. pp. 43-60.

Solomon, M., Peters, K.J. and Azage, T., 2004. Effect of supplementation with foilages of selected multipurpose trees, their mixtures or wheat bran on feed intake, plasma enzyme activities, live weight and scrotal circumference gain in Menz sheep. *Livestock production science*. **89**: 253-264.

Spencer, D., Higgins, T.J.V., Freer, M., Dove, H. and Coombe, J.B., 1988. Monitoring the fate of dietary proteins in rumen fluid using gel electrophoresis. *Br J Nutr*. **60**: 241–247.

Szyszkowski, M., Ter Menlen, U. and El-Harith, E.A., 1983. The possibilities of safe application of *Leucaena leucocephala* in the diets of productive livestock. *Leucaena Res. Rep.* **4**:13-14.

Tefera, S.; Mlambo, V.; Dlamini, B. J.; Dlamini, A. M.; Koralagama, K. D. N. and Mould, F. L., 2008: Chemical composition and in vitro ruminal fermentation of common tree forages in the semi-arid rangelands of Swazi-land. *Animal Feed Science and Technology*. **142**: 99-110.

Tessema, Z., Baars, R.M.T. and Alemu, Y., 2003. Effect of plant height at cutting and fertiliser on growth of Napier grass (*Pennisetum purpureum* Schumach.). *Tropical Science*. **43**: 57-61.

Thadei, C., Kimambo, A.E. and Mushi, D.E., 2001. Relationship between tannins concentration in plant materials and their effects on protein degradation in the rumen. Department of Animal Science and Production, Denmark. *TSAP Proceedings*. **28**.

Thomas, C., 2004. Feed into Milk: A New Applied Feeding System for Dairy Cows: An Advisory Manual. Nottingham, UK: Nottingham University Press.

Tilley, J.M.A. and Terry; R.A., 1963. A two-stage technique for the in vitro digestion of forage crops. *J Br Grassland Soc.* **18**: 104-111.

Topps, J. H., 1992: Potential, composition and use of legume shrubs and trees as fodder for livestock in the tropics – review. *Journal of Agricultural Science*. **118**: 1-18.

Tylutki, T. P.; Fox, D. G.; Durbal, V. M.; Tedeschi, L. O.; Russell, J. B.; Van Amburgh, M. E.; Overton, T. R.; Chase, L. E. and Pell, A. N., 2008: Cornell net carbohydrate and protein system: a model for precision feeding of dairy cattle. *Animal Feed Science and Technology*. **143**: 174-202.

Van Hellen, R. W. and Ellis, W. C., 1977. Sample container porosities for rumen in situ studies. *J. Anita. Sci.* **44**: 141.

Van Keuren, R.W. and Heinemann, W.W., 1962. Study of the nylon bag technique for in vivo estimation of forage digestibility. *J Anim Sci.* **21**: 340-345.

Van Raay, H.G.T. and De Leeuw, P.N., 1970. The importance of crop residues as a fodder: A resource analysis in Katsina Province, Nigeria. **61**: 137-147.

Van Raay, H.G.T. and De Leeuw, P.N., 1974. Fodder Resources and Grazing Management in a Savanna Environment: An Ecosystems Approach. Occasional Paper No. 5, Institute of Social Studies, the Hague, the Netherlands.

Van Soest, P.J., 1982. Nutritional Ecology of the Ruminant. O & B Books, Inc. Corvallis, Oregon. USA. pp. 76-78.

Van Soest, P. J., 1994: Nutritional Ecology of the Ruminant, 2nd edn. Comstock Publishing Associates/Cornell University Press, Ithaca, NY, USA.

Van Soest, P.J., Robertson, J.B. and Lewis, B.A., 1991. Methods of dietary fibre, neutral detergent fibre and non-starch monosaccharides in relation to animal nutrition. *Journal of Dairy Science.* **74**: 3583-3597.

Vanzant, E. S., Cochran, R. C. and Titgemeyer, E. C. r., 1998. Standardization of in situ techniques for ruminant feedstuff evaluation. *J. Anim. Sci.* **76**: 2717-2729.

Verma, D.N., 2006. A Textbook of Animal Nutrition. Kalyani Publishers, New Delhi. pp. 126-132.

Villalba, J. J. and Provenza, F. D., 1996. Preference for flavored wheat straw by lambs conditioned with intraruminal administrations of sodium propionate. *J. Anim. Sci.* **74**: 2362-2368.

Villalba, J. J. and Provenza, F. D., 1999. Nutrient-specific preferences by lambs conditioned with intraruminal infusions of starch, casein, and water. *J. Anim. Sci.* **77**: 378-387.

Vranic, M., Mladen, K., Kresimir, B., Josip, L., Goran, P., Hrvoje, K. and Ivana, M., 2009. Maize silage supplementation to lower quality grass silage improves the intake, apparent digestibility and N retention. *Mljekarstvo.* **59(4)**: 302-310.

Waghorn, G. C. and Shelton, I. D., 1997. Effect of condensed tannins in *Lotus corniculatus* on the nutritive value of pasture for sheep. *J. Agric. Sci. Camb.* **128**: 365-372.

Web site: [Http://en.wikipedia.org/wiki/Holeta_Genet](http://en.wikipedia.org/wiki/Holeta_Genet)

Web site: [Http://en.wikipedia.org/wiki/Jimma](http://en.wikipedia.org/wiki/Jimma)

Web site: [Http://en.wikipedia.org/wiki/Kersa_Jimma_zone](http://en.wikipedia.org/wiki/Kersa_Jimma_zone)

Welch, J.G., 1986. Physical parameters of fiber affecting passage from the rumen. *J. Dairy Sci.* **69**: 2750-2754.

Whetton, M.; Rossiter, J. T. and Wood, C. D., 1997: Nutritive evaluation of nitrogenous fractions in leaves of *Gliricidia sepium* and *Calliandra calothyrsus* in relation to tannin content and protein degradation by rumen microbes. *Journal of Agricultural and Food Chemistry.* **45**: 3570-3576.

Woldemeskel, M.; Tegegne, A.; Umunna, N. N.; Kaitho, R. J. And Tamminga, S., 2001. Effects of *Leucaena pallida* and *Sesbania sesban* supplementation on testicular histology of tropical sheep and goats. *Anim. Repr. Sci.* **67(3/4)**: 253-265.

Wolmer, W., 1997. Crop–livestock integration: The dynamics of intensification in contrasting agro ecological zones: A review. Institute of Development Studies, University of Sussex, UK. pp 30.

Wood, C.D, Grillet, C., Resales, M. and Green, S., 1995. Relationships between *in vitro* gas production characteristics and composition of tree leaf fodders from Bolivia, West Africa and Colombia. *Abstract Animal Science.* **60**: 541.

World Bank., 1989. Sub-Saharan Africa: From Crisis to Sustainable Growth. Washington DC, USA.

Yisehak, K., Becker, A., Rothman, J.M., Dierenfeld E.S., Marescau B., Bosch G., W. and Janssens G.P.J., 2012. Amino acid profile of salivary proteins and plasmatic trace Hendriks mineral response to dietary condensed tannins in free-ranging zebu cattle (*Bos indicus*) as a marker of habitat degradation. *J. Livestock Science.* **144** (3): 275-280.

Yisehak, K., Belay, D. and Janssens, G.P.J., 2010. Indigenous fodder trees and shrubs: Nutritional and ecological approaches. Proceedings of 14th ESVCN Congress, Zurich, Switzerland. pp.121-121.

Yisehak, K., Belay, D., Solomon, D. and Janssens, G.P.J., 2009. Adaptation of cattle to tannin rich diet. Proceedings of 13th ESVCN Congress, Sardinia, Italia. pp.110-110.

Yisehak, K., Solomon, M. and Tadele, M., 2011. Contribution of *Moringa (Moringa stenopetala, Bac.)*, a highly nutritious vegetable tree, for food security in South Ethiopia: A review. *Asian J. Applied Sci.* **4(5)**: 477-488.

7. APPENDICES

7.1. Appendix. Chemical Composition and *In Sacco* degradability Tables

Appendix Table 1. The least square means for chemical compositions (%) of plant species

Plant species	PP	Chemical composition, mean							
		DM	CA	CP	EE	NDF	CF	NFC	CT
<i>E. capensis</i>		96.84 ^a	7.91 ^c	9.76 ^d	5.06 ^a	37.96 ^c	19.10 ^{ab}	39.30 ^a	8.00
<i>F. sycomorus</i>		92.66 ^c	11.49 ^a	12.42 ^c	1.57 ^b	61.01 ^a	15.40 ^b	13.51 ^{bc}	8.49
<i>M. lanceolata</i>	leaf	95.10 ^b	7.15 ^d	17.06 ^a	5.17 ^a	61.33 ^a	21.45 ^a	8.71 ^c	13.65
<i>R. glutinosa</i>		96.86 ^a	9.29 ^b	16.04 ^b	4.70 ^a	49.31 ^b	12.20 ^c	20.66 ^b	16.72
	SE	0.06	0.17	0.27	0.16	2.46	1.49	2.41	1.66
	P	***	***	***	***	***	**	**	NS
<i>E. capensis</i>		94.11 ^c	5.43 ^d	11.07	31.00 ^a	35.90 ^d	20.25	17.28 ^a	7.27
<i>F. sycomorus</i>	fruit	95.56 ^b	9.70 ^a	10.81	5.97 ^c	66.71 ^b	27.5	9.07 ^{bc}	8.17
<i>M. lanceolata</i>		96.5 ^a	7.45 ^b	7.97	8.20 ^b	62.00 ^c	22.20	14.38 ^b	7.20
<i>R. glutinosa</i>		92.43 ^d	5.57 ^c	11.21	5.33 ^d	68.9 ^a	22.30	5.84 ^c	8.59
	SE	0.30	0.30	2.04	0.18	1.26	2.41	2.39	0.86
	P	***	***	NS	***	***	NS	*	NS

PP, plant part; DM, dry matter; CA, crude ash; CP, crude protein; EE, ether extract, NDF, neutral detergent fiber; CF, crude fiber; NFC, non-fibre carbohydrates; CT, condensed tannin; ^{a, b, c, d} Least square means with the different letters within the column are significantly different ($P < 0.05$); * $P < 0.05$; *** $P < 0.001$; NS, non-significant ($P > 0.05$).

Appendix Table 2. The Least square means of *in sacco* ruminal dry matter disappearance (%) of the plant species

Plant species	PP	Incubation time (h), mean (%)							
		0	6	12	24	48	72	96	
<i>E. capensis</i>		29.43 ^a	32.12 ^a	59.47 ^a	73.11 ^a	82.49 ^a	81.79 ^a	83.38 ^a	
<i>F. sycomorus</i>	Leaf	6.32 ^d	15.82 ^b	26.09 ^c	22.53 ^b	61.07 ^b	59.52 ^b	69.89 ^c	
<i>M. lanceolata</i>		23.69 ^b	32.60 ^a	35.59 ^b	64.99 ^a	65.72 ^b	81.46 ^a	77.53 ^b	
<i>R. glutinosa</i>		19.28 ^c	16.87 ^b	17.99 ^c	23.72 ^b	38.20 ^c	47.07 ^c	47.64 ^d	
	SE	0.68	1.88	3.13	3.87	4.71	2.79	1.47	
	P	***	***	***	***	***	***	***	
<i>E. capensis</i>		39.28 ^a	43.34 ^a	60.01 ^a	62.02 ^a	78.95 ^a	75.94 ^a	84.85 ^a	
<i>F. sycomorus</i>	Fruit	20.81 ^b	23.29 ^b	34.20 ^b	43.17 ^b	58.90 ^b	62.34 ^b	69.89 ^b	
<i>M. lanceolata</i>		10.32 ^d	15.14 ^c	19.03 ^c	28.64 ^c	40.11 ^c	54.04 ^c	46.08 ^c	
<i>R. glutinosa</i>		17.60 ^c	17.91 ^b	20.30 ^c	22.16 ^c	26.46 ^d	34.16 ^d	21.65 ^d	
	SE	0.45	2.12	1.49	3.37	2.44	2.79	1.57	
	P	***	***	***	***	***	***	***	

PP, plant species; ^{a, b, c, d} Least square means within the same column with different superscripts are significantly different ($P < 0.05$); SE, Standard error of the mean; *** $P < 0.001$.

Appendix Table 3. *In sacco* dry matter degradability characteristics for the leaves and fruits of the plant species

Plant species	Degradability parameters (leaf)					Degradability parameters (fruit)				
	a	b	PD	ED	c	A	b	PD	ED	C
<i>E. capensis</i>	-16.2 ^d	98.37 ^a	82.17 ^b	61.83 ^a	0.12 ^a	35.51 ^a	47.11 ^b	82.62 ^a	63.18 ^a	0.04 ^b
<i>F. sycomorus</i>	7.37 ^c	77.82 ^b	85.19 ^a	35.78 ^c	0.02 ^c	15.49 ^b	56.15 ^a	71.64 ^b	43.28 ^b	0.03 ^c
<i>M. lanceolata</i>	14.55 ^a	64.91 ^c	79.47 ^c	53.76 ^b	0.05 ^b	5.78 ^d	46.59 ^c	52.36 ^c	29.22 ^c	0.03 ^c
<i>R. glutinosa</i>	11.23 ^b	52.76 ^d	63.99 ^d	27.86 ^d	0.01 ^d	13.92 ^c	13.69 ^d	27.61 ^d	22.57 ^d	0.05 ^a
SE	0.17	0.03	0.12	0.05	0.001	0.007	0.01	0.01	0.01	0.004
P	***	***	***	***	***	***	***	***	***	***

a, soluble fraction; *b*, slowly degradable fraction; *PD*, potential degradability; *ED*, effective degradability; *c*, rate of degradation; ^{a, b, c, d} Least square means within the same column with different superscripts are significantly different ($P < 0.05$); *SE*, Standard error of the mean; *** $P < 0.001$

Appendix Table 4. Least square means for *in sacco* ruminal organic matter disappearance (%) of leaf and fruit of tannin rich trees

Plant species	PP	Incubation time (h), mean (%)						
		0	6	12	24	48	72	96
<i>E. capensis</i>		29.43 ^a	37.55 ^a	61.05 ^a	73.28 ^a	82.48 ^a	81.79 ^a	83.38 ^a
<i>F. sycomorus</i>	Leaf	6.32 ^d	22.63 ^c	26.45 ^c	23.57 ^b	62.64 ^b	61.05 ^b	69.89 ^c
<i>M. lanceolata</i>		23.70 ^b	31.21 ^{ab}	36.05 ^b	65.91 ^a	72.30 ^{ab}	81.69 ^a	77.60 ^b
<i>R. glutinosa</i>		19.28 ^c	17.44 ^c	20.63 ^c	26.26 ^b	42.02 ^c	47.27 ^c	48.98 ^d
	SE	0.68	3.49	3.03	3.54	3.46	2.36	1.43
	P	***	***	***	***	***	***	***
<i>E. capensis</i>		39.28 ^a	43.34 ^a	60.68 ^a	64.81 ^a	81.34 ^a	80.29 ^a	84.99 ^a
<i>F. sycomorus</i>	Fruit	20.81 ^b	21.27 ^b	34.96 ^b	45.43 ^b	53.40 ^b	64.88 ^b	69.36 ^b
<i>M. lanceolata</i>		10.32 ^d	14.85 ^b	19.03 ^c	29.73 ^c	47.86 ^b	54.61 ^c	45.63 ^c
<i>R. glutinosa</i>		17.60 ^c	17.91 ^b	20.30 ^c	24.39 ^c	27.73 ^c	38.25 ^d	31.09 ^d
	SE	0.45	2.44	1.60	2.91	2.31	1.73	3.50
	P	***	***	***	***	***	***	***

PP, plant part; ^{a, b, c, d} Least square means within the same column with different superscripts are significantly different ($P < 0.05$); *** $p < 0.001$; *SE*, Standard error of the mean

Appendix Table 5. *In sacco* organic matter degradability characteristics of leaves and fruits of selected browses species

Plant species	Degradability parameters (<i>leaf</i>)					Degradability parameters (<i>fruit</i>)				
	a	b	PD	ED	c	a	b	PD	ED	C
<i>E. capensis</i>	10.69 ^b	45.21 ^d	55.90 ^d	29.61 ^d	0.02 ^c	14.22 ^c	21.38 ^d	35.60 ^d	24.58 ^d	0.03 ^c
<i>F. sycomorus</i>	12.63 ^a	77.27 ^b	89.89 ^a	38.11 ^c	0.02 ^c	15.26 ^b	56.40 ^a	71.66 ^b	42.84 ^b	0.03 ^c
<i>M. lanceolata</i>	8.11 ^c	71.98 ^c	80.09 ^c	54.55 ^b	0.05 ^b	2.06 ^d	50.28 ^c	52.34 ^c	30.59 ^c	0.04 ^b
<i>R. glutinosa</i>	-1.02 ^d	83.35 ^a	82.33 ^b	63.97 ^a	0.11 ^a	31.84 ^a	52.02 ^b	83.86 ^a	64.65 ^a	0.05 ^a
SE	0.02	0.009	0.08	0.04	0.002	0.01	0.06	0.05	0.007	0.001
P	***	***	***	***	***		***	***	***	***

a, soluble fraction; *b*, slowly degradable fraction; *PD*, potential degradability; *ED*, effective degradability; *c*, rate of degradation; ^{a, b, c, d}Least square means within the same column with different superscripts are significantly different ($P < 0.05$); *** $p < 0.001$; *SE*, Standard error of the mean